

# Exercise and Immunity: A Correlated Mechanism

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## ABSTRACT

Sedentary lifestyles now become an incredible worldwide problem with the advancement of technologies over the past several decades. Physical inactivity has progressively increased now-a-days and potentially contributes the risk of numerous diseases/disorders. The initial body defensive mechanisms contribute the protective responses, effective against a diverse variety of threats. Regular physical activity, fitness and exercise are critically important for the good health and may have prevalence over immunological system. Prolonged bouts of exercise are correlated with depressed immune system to pick up opportunistic infections. Moderate exercise whereas elicit beneficial outcomes in both prevention and rehabilitation of many diseases. Nevertheless the cascades of exercise-induced cellular-molecular-signaling pathway may trigger antigen-receptor recognition gateway. Exercise and immunity henceforth, is a correlated mixed-message mechanism and has important potential implications on public health.

**Key Words:** Physical-activity; NK cells; ROS; SIgA; Myokine; Autophagy

## INTRODUCTION

Physical activity is the bodily movement, constructed by skeletal muscles that outcomes in calorie expenditure. <sup>[1]</sup> Henceforth, physical activity is multidimensional, portrayed by frequency, duration, intensity and type of activity. <sup>[1,2]</sup>

Exercise (or exercise-training), a subcategory of physical activity, is planned, structured, repetitive and intentional bodily movements intended to improve or maintain one or more components of physical fitness. <sup>[2-4]</sup>

An important difference between physical activity and fitness is the intra-individual day-to-day variability. <sup>[1]</sup> Physical activity varies indubitably on a daily basis, whereas fitness remains relatively static and taking time to change.

Physical activity is an input to countless surveillance and epidemiological studies, considering trends and connections with disease. Public health initiatives

intended at escalating physical activity, resort to monitor the measurement of physical activity. <sup>[1]</sup> Exercise, a subcategory of physical activity, relates to immunity is a combined memorandum. <sup>[5]</sup> Regular moderate-intensity exercise diminishes the risk of infection, <sup>[5,6]</sup> whereas prolonged or over-trained exercise is associated with immunosuppression. <sup>[7-9]</sup> Exercise may increase the production of macrophages that assaulting bacteria, associated with upper respiratory tract diseases (URTD). <sup>[5,7]</sup> Cross-sectional and longitudinal data recommend that regular moderate-intensity exercise uphold a reduced risk of self-reported respiratory symptoms. <sup>[7,10-12]</sup> Temporary rise in body temperature during exercise may inhibit bacterial growth as well as sluggish stress-related hormone. <sup>[5,13]</sup> However, over-exercise emerges to negatively affect immunity. Quinn <sup>[14]</sup> accounted that >90 minutes of high-

intensity exercise (marathons, triathlons, endurance races) makes a person more susceptible to illness for up to 72 hours after working out. During prolonged or high-intensity exercise, the body produces stress-hormones (cortisol and adrenaline), that raise blood pressure, elevate cholesterol levels and suppress the immune system. [5,14] Intense exercise before or during viral infection has been associated with greater morbidity and mortality. [7,15,16] It is well established by Nieman [17] “J-shaped” hypothesis, representing the relation between exercise and susceptibility to infection. This model suggests that immune function enhanced above sedentary levels with moderate activity while excessive amounts of prolonged, high-intensity exercise may impair immune function. Nevertheless, strenuous physical exercise executes increased-vulnerability to immunological mechanism is well defined by “open window” theory. [9,18-20] “Open window” for pathogen entrance may alter immunity via the risk of clinical infection after excessive exercise which can last between 3 to 72 hours. According to the International Society of Exercise and Immunology (ISEI), the immune dysfunction remarkably observed due to continuous and prolonged (> 1.5h) exercise and performed at an intensity ranging from moderate to high. [8]

The purpose of this article is to summarize current literatures regarding exercise and immunity as well as to provide a platform for further investigation into the mechanisms and reconciling the protective effect of exercise. The study narrates how an exercise stimulus may act upon the inter-dependent network to orchestrate the beneficial or reverse effects on pathophysiology as well as immunity.

#### **Exercise and Immunological Features**

Exercise of moderate intensity stimulates parameters related to cellular immunity to decrease the risk of infection, while high-intensity exercise may promote a reduction of these same parameters, increasing the risk of infectious diseases.

[21,22] The line of defense engrosses white blood cells (leukocytes) that travel through the bloodstream and into tissues, searching for and attacking microorganisms and other invaders. [23] Nielsen, [20] Pederson & Hoffman-Goetz, [21] McCarthy & Dale [24] and Fry et al. [25] stated that the alteration in leukocyte numbers in circulating blood is the most studied aspects of exercise and immune system. During exercise the chief source of circulatory neutrophils are bone marrows, spleen, lymph nodes and gut. [20,26,27] Fry et al., [25] Nieman et al. [28] and Peake [29] scrutinized that intensity of exercise, duration and/or the fitness level can cooperate with the degree of leukocytosis occurring.

Leukocytes has two modes of actions in term of internal-physical defense- i) innate immunity and ii) acquired immunity.

#### **Innate Immunity**

Innate immunity, the first line of defense, responds to invaders instantly without learning to recognize them. It consists of anatomic, physiological and chemical barriers such as skin, body temperature, pH, complement systems. [20,22] Natural killer (NK) cells, neutrophils, macrophages, as well as microbicide molecules such as the nitric oxide (NO) and free radicals are the participants of innate immunity.

NK cells, one of the first innate lymphoid cell populations can display cytotoxicity by producing cytokine, IF $\gamma$ . [30] NK cells present remarkable sensitivity to the stress induced by physical exercise, which promotes their redistribution from the peripheral blood to other tissues. [22] Timmons & Cieslak [30] suggested that the NK cells may be a potential link between regular physical activity and general health status. Mobilization of peripheral circulation of NK cells may transpire via mechanisms including, stress caused by a substantial increase in the peripheral blood flow and decreased expression of adhesion molecules like catecholamine, [31] by physical exercise. [32] Jones & Hoyne [33] observed a transient

increase of NK cells with a rapid egress, in the acute post-exercise phase 1 hour following completion of exercise. Gannon et al. [34] however, reported that the concentration of circulating NK cells may return to the pre-exercise level, or even become lower during excessive prolonged exercise (>3h).

Leukocytes circulate through the body and work in a coordinated manner to monitor the body for invaders. [20] Mainly two types of leukocytes (phagocytes and lymphocytes) engage with immunity. Neutrophils are phagocytes, involved in many of inflammatory processes of muscle fiber to raise calcium release and synthesis of pro-inflammatory cytokines (tumor-necrosis-factor-TNF- $\alpha$  and inter-leukine-IL-1 $\beta$ ), promoted by exercise, playing an important role in the innate immune response. [22] Walsh [35] evaluated that intense physical exercise endorses degranulation of neutrophils to enhance the concentration of enzymes - myeloperoxidase (MPO), which acts as a marker of neutrophil migration into the muscle and of the degranulation in the serum. Morozov et al [36] concluded with an experiment that a single session of exhaustive exercise produces significant MPO in untrained group compared to the trained group, suggesting a possible protective effect from training in the muscle tissue.

Monocytes are effective phagocytes, responsible to differentiate into macrophages. Macrophages are liable to give innate immunity by phagocytosis, microbial killing and antitumor activity [37] but also predominant in triggering atherosclerosis. [20] Both moderate and exhaustive exercise enhances a variety of peritoneal macrophage capacities, including chemotaxis [38,39] adherence, [40] phagocytosis [41,42] and antitumor [43] activity. Potent effects of exercise on macrophage function may be interceded by exercise-induced alterations in the sympathetic-nervous-system or hypothalamic-pituitary-axis via

neuroendocrine hormones in a local manner. [37,44] Kohut et al. [45] found that the exercise-induced suppression in intrinsic alveolar macrophage antiviral resistance can be abrogated by propranolol, a  $\beta$ -adrenergic antagonist. According to Woods et al. [46] monocyte numbers transiently increase (50-100%) in peripheral blood in response to acute exercise. Grabieli et al. [47] demonstrated that exercise intensity or duration-dependent changes in subpopulations of monocytes may migrate out of the vasculature following long-duration exercise. Woods et al. [37] evaluated that both moderate and exhaustive treadmill running over periods of 3-7 days increases the antitumor activity of thioglycolate (TG)-elicited or *Propionibacterium acnes*-activated peritoneal macrophages due to enhance production of TNF $\alpha$  and NO respectively though other peritoneal macrophage functions like antigen presentation may be suppressed by exercise.

This increase in molecular oxygen ( $O_2$ ) consumption combined with the activation of specific metabolic pathways during and after exercise can result in free radicals or reactive-oxygen-species (ROS) formation. [48-51] ROS can be fabricated by different mechanisms during physical exercise, including the partial diminution of oxygen in the mitochondria (electron transport chain) and the inflammatory process. [51-54] ROS, produced by metabolically oxidative process are required for the activation of immune system. [55] ROS also can induce pro-inflammatory cytokines. [56] The intracellular ROS serve mainly for host defense against infectious agents, redox-sensitive signal transduction and other cellular processes, while the extracellular release of ROS may damage surrounding tissues, potentially promoting inflammatory processes. [56-58] Jin et al. [59] accounted that the elevated oxidative and physical stress reflected by the level of intracellular ROS and cortisol respectively, may contribute to immunosuppression. In addition, low levels of inflammatory markers observed in the elderly who

frequently exercise. [60] Acute exercise generates excessive ROS causing damage in the body, while regular exercise results in bodily adaptations leading to resistance against oxidative damage via antioxidant pathways. [61,62] Yavari et al. [61] reported that mitochondrial ROS generated during regular exercise are necessary for the activation of primary signaling pathways associated with muscle adaptation. Nuclear factor erythroid 2-related factor (Nrf2), a redox-sensing transcription factor, is the primary regulator of antioxidants as well as other cytoprotective cofactors that are responsible for the enhanced antioxidant defense system. [62-64]

### **Relation with Mucosal Immunity**

The mucosal immune system protects mucosal surfaces of the respiratory tract, nasal passages and intestine. Saliva is the most commonly used secretion for measurement of secretory antibodies in the assessment of mucosal immune status. [65] The effect of acute exercise on mucosal immunity is mainly focused on changes in the secretion of immunoglobulin A (SIgA) as determined in saliva. [66,67] Secretory IgM (SIgM) antibodies contribute to a lesser extent in the normal adult but play a significant role in mucosal defense in the neonate and also in IgA-deficiency states. [65,68] Individuals with selective IgA deficiency suffer from upper respiratory tract infection (URTI). [69-71] Habitual exercise at an intense level can cause suppression of mucosal immune parameters, while moderate exercise may have positive effects. [65,72] According to Gleeson & Pyne [65] the diminished mucosal immunoglobulins after exercise do not appear to be the upshot of the mucosal plasma cells depletion but recovery to pre-exercise levels usually occurs within 24 hours, though after high-intensity exercise the levels may remain suppressed for longer period leading to increased-risk of infection. [8] Nieman (2000) [13] and Müns et al. [73] reported that nasal mucociliary transit time is significantly prolonged by abnormally functioning ciliated cells after a marathon

race for several days. This situation impairs in nasal neutrophil function and nasal/salivary IgA secretion rates, directing the suppressed host protection in the upper airway passages.

### **Adaptive Immunity**

Lymphocytes (B cells and T cells) represent adaptive or acquired immunity to learn how to efficiently encounter an invader and remember the specific invader. [23] T lymphocytes including T-helper-cell-type-1 (Th1, with surface protein cluster of differentiation CD4+), T-helper-cell-type-2 (Th2, CD4+) and cytotoxic-T-cell (Tc, with surface protein cluster of differentiation CD8+) recognize invaders via antigen-presenting cells (APCs) like macrophages and dendritic cells for recognition, activation, mobilization, regulation and resolution to represent cellular adaptive immunity. [74] The ratio of CD4+: CD8 declines throughout exercise, reflecting a more remarkable enhancement of Tc cells [21] due to increase in numbers of NK cells more than any other T cell subpopulation. [30] Prolonged and extenuating aerobic exercises reduce the expression of Toll-like receptors (TLRs) in macrophages, compromising the presentation of antigens for the Th1 inflammatory response. [22] The Th1 anti-inflammatory effect evades the usual tissue damage and reduces the risk of chronic inflammatory diseases but raises the susceptibility to infections by intracellular microorganisms. [75]

B lymphocytes and their products antibodies and cytokines represent humoral adaptive immunity via neutralization, agglutination, precipitation, complement activation and lysis. [22,76] Acute Exercise induces muscle cell injury due to a sequential release of the pro-inflammatory cytokines TNF- $\alpha$ , IL-1b and IL-6 followed by anti-inflammatory cytokines. [18,77,78] Endurance exercise (marathon running) associated with muscle soreness, induces a greater inflammatory cytokine response than other modes like cycling or rowing. [18] Bruunsgaard et al. [79] evaluated that the delayed-type hypersensitivity (DTH)



reaction was suppressed in triathletes (60%), two days after competing in a half iron man triathlon (mean time 6.5 h).

Contracting muscle can produce and release a cytokine named myokine (IL-6), exerting autocrine, paracrine or endocrine effects. [80] Myokines provide a conceptual basis to explain how muscles communicate to other organs. [81] Pedersen & Febbraio [80] appraised that intramuscular IL-6 expression is modified by signaling network involving crosstalk between the Ca<sup>2+</sup>/nuclear factor of activated T-cells (NFAT) and lycopogen/p38 mitogen-activated protein kinase (MAPK) pathways. Consequently, macrophage-produced IL-6 leads to an inflammatory response, whereas muscle cells fabricate and release IL-6 without activating classical pro-inflammatory pathways.

#### **Signaling Pathways Involved In Exercise Induced Immune Response**

The ligand-receptor molecular interactions trigger biochemical cascade signaling pathways, resulting in the production of proteins, cytokines and the expression and proliferation of receptors. [22] The antigen-receptor aggregation leads to an activation of tyrosine kinase proteins by phosphorylation, associated with the receptors in the cellular membrane. [82] IL-6 activates the signaling pathway of the Janus-activated-kinase (JAK) as well as signal transducer and activator of transcription (STAT) protein complex by phosphorylation and activation of the mitogen-activated-protein-kinase (MAPK)/extracellular-signal-regulated-kinase (ERK) signaling cascade with interference by adenosine-mono-phosphate-kinase (AMPK) and phosphoinositol-3-kinase - protein-kinase-B (PI3K-AKT) signaling. [80] In the immunological system, mammalian target of rapamycin (mTOR; serine/threonine kinase) protein signaling is triggered by the antigens ligation to their specific receptors in T and B cells or to TLR and by the ligation of interleukins to their receptors. [22,83,84]

Exercise may cause the production of growth factors and cytokines. [22]

Exercise training strongly alters physical forces, exerting significant physiologic effects on endothelial and smooth muscle gene expression and function. [85,86] Exercise induced mechanotransduction mechanisms trigger many intracellular pathways like MAPK and sequential phosphorylation activating transcription factors and gene expression. [87,88] Thompson et al. [84] reported that exercise promotes cellular stress and DNA damage that may inhibit the type of mTOR activity, leading to a pro-inflammatory effect in phagocytic cells to produce cytokines (IL-6, IL-12 and IL-23) and decrease the production of anti-inflammatory cytokines ( IL-10). This phenomenon triggers the cellular immunity. [89]

#### **Exercise Induced Beneficial Effect of Cellular System**

Exercise has numerous health benefits, including lifespan expansion and protection against cardiovascular diseases, diabetes, cancer and neurodegenerative diseases. [90,91] Many of these health benefits overlap with cellular programmed mechanism [92,93] triggering through various signaling pathways of cellular proteins and/or protein kinases. Autophagy (programmed cell death type II; PCDII), a high-capacity process, encloses selective elimination of vital organelles and/or proteins, instigating mechanisms of cytoprotection and homeostasis in biological systems under normal physiological and stress conditions. [94] Autophagy is an evolutionarily conserved adaptive ubiquitous cellular process governed by dynamic catabolic biochemical mechanisms to generate autophagosomes by engulfing intracellular components and ultimately fuse with lysosomes. [94-97]

He et al. [91] reported that acute exercise (30 min) can sufficiently induce autophagosome formation in skeletal and cardiac muscle, reaching a plateau phase after 80 min of exertion. Accordingly, by using several markers of autophagy, He et al. [91] also evaluated that there is a possible role of autophagy in metabolic regulation

during exercise in skeletal muscle, heart, liver, pancreatic  $\beta$  cells and adipose tissue. A single bout of long endurance exercise increases autophagic flux via the ubiquitin-proteasome and the autophagolysosome systems, associated with mTOR signaling. [98,99] Chronic exercise affects the basal level of the autophagy by increasing the transcription of several autophagic proteins/genes. Autophagy modulation by exercise is linked to the nutritional status, are also modulated by amino acid and glucose availability. [99,100] Exercise may elicit metabolic stress, oxidative stress, calcium imbalance and general disturbances in cellular homeostasis [101-103] which may contribute to autophagic activation. [91,97,104] Grumati et al. [105] suggested that the functional autophagy is required for the proper response to acute and chronic exercise. Moreover, life-long exercise, in mishmash with caloric restriction, improved the decline in autophagic-proteins with aging and dampened the age-related increase in oxidative damage. [106]

## CONCLUSIONS

Life has changed dramatically over the last century. People have become less active, adopting more sedentary habits. A sedentary lifestyle potentially contributes the serious implications on human-health and produce chronic diseases such as cardiovascular diseases, obesity, type-2-diabetes and metabolic syndromes. In this scenario, regular practice of physical exercise should be beneficial to health by altering the metabolic state and also the immune system. Overall this review summarized that moderate exercise may have predominance over cellular and humoral immunity whereas high intensity exercise raises the concentration of anti-inflammatory cytokines, leading to cause infection by intracellular microorganisms. Exercise induced mechanotransduction mechanism triggers many intracellular pathways like signaling-cascades of MAPK/ERK, AMPK, PI3-AKT, mTOR, associated with antigen-receptor recognition

gateway of immunology. Muscle can release important factors via activation of autophagy which may foster the pleiotropic health benefits of exercise. The current state of knowledge hereafter may shed a light on the alteration of immune response depending on the types of exercise.

## Competing Interests

The authors declare that they have no competing interests.

## REFERENCES

1. Warren JM, Ekelund U, Besson H, Mezzani A, Geladas N, Vanhees L. Assessment of physical activity - a review of methodologies with reference to epidemiological research: a report of the exercise physiology section of the European Association of Cardiovascular Prevention and Rehabilitation. *European Journal of Cardiovascular Prevention and Rehabilitation* 2010; 17(2):127–139.
2. Caspersen CJ, Powell K., Christenson GM. Physical activity, exercise and physical fitness: definitions and distinctions for health-related research. *Public Health Reports* 1985; 100:126–131.
3. Howley ET. Type of activity: resistance, aerobic and leisure versus occupational physical activity. *Medicine and Science in Sports and Exercise* 2001; 33:S364–S369.
4. Gummelt D. Physical Activity vs. Exercise: What's the Difference? June 3, 2015; Retrieved from: <https://www.acefitness.org/education-and-resources/lifestyle/blog/5460/physical-activity-vs-exercise-what-s-the-difference>.
5. Brown J. How Exercise Affects Immunity. EXOS Knowledge. March 11, 2013; Retrieved from: <http://www.coreperformance.com/knowledge/wellness/how-exercise-affects-immunity.html>.
6. Nieman DC, Henson DA, Austin MD, Sha W. Upper respiratory tract infection is reduced in physically fit and active adults. *British Journal of Sports Medicine* 2011; 45(12):987–992.
7. Martin SA, Pence BD, Woods JA. Exercise and respiratory tract viral infections. *Exercise and Sports Sciences Reviews* 2009; 37(4):157–164.

8. Walsh NP, Gleeson M, Pyne DB, Nieman DC, Dhabhar FS, Shephard RJ, Oliver SJ, Bermon S, Kajeniene A. Position statement part two: maintaining immune health. *Exercise Immunology Review* 2011; 17:64–103.
9. Gleeson M. Effects of exercise on immune function. *Sports Science Exchange* 2015; 28(151):1–6.
10. Nieman DC, Johanssen LM, Lee JW. Infectious episodes in runners before and after a road race. *Journal of Sports Medicine & Physical Fitness* 1989; 29(3):289–96.
11. Kostka T, Berthouze SE, Lacour J, Bonnefoy M. The symptomatology of upper respiratory tract infections and exercise in elderly people. *Medicine and Science in Sports and Exercise* 2000; 32(1):46–51.
12. Matthews CE, Ockene IS, Freedson PS, Rosal MC, Merriam PA, Hebert JR. Moderate to vigorous physical activity and risk of upper-respiratory tract infection. *Medicine and Science in Sports and Exercise* 2002; 34(8):1242–1248.
13. Nieman DC. Exercise effects on systemic immunity. *Immunology and Cell Biology* 2000; 78:496–501.
14. Quinn E. Health and safety: Too much exercise and decreased immunity. Can too much exercise decrease your immunity and make you sick? September 23, 2017; Retrieved from: <https://www.verywellfit.com/exercise-and-immunity-3120439>.
15. Heath GW, Ford ES, Craven TE, Macera CA, Jackson KL, Pate RR. Exercise and the incidence of upper respiratory tract infections. *Medicine and Science in Sports and Exercise* 1991; 23(2):152–157.
16. Ekblom B, Ekblom O, Malm C. Infectious episodes before and after a marathon race. *Scandinavian Journal of Medical Science in Sports* 2006; 16(4):287–293.
17. Nieman DC. Exercise, infection and immunity. *International Journal of Sports Medicine* 1994; 15(13):S131–S141.
18. Nieman DC, Nehlsen-Cannarella SL, Fagoaga OR, Henson DA, Utter A, Davis JM, Williams F, Butterworth DE. Influence of mode and carbohydrate on the cytokine response to heavy exertion. *Medicine and Science in Sports and Exercise* 1998; 30:671–678.
19. Shephard RJ. Sepsis and mechanisms of inflammatory response: is exercise a good model? *British Journal of Sports Medicine* 2001; 35(4):223–230.
20. Nielsen HG. Sports and exercise medicine: Exercise and immunity: 2013.p,122–140 INTECH. <http://dx.doi.org/10.5772/54681>.
21. Pedersen BK, Hoffman-Goetz L. Exercise and the immune system: regulation integration and adaption. *Physiological Reviews* 2000; 80:1055–1081.
22. Terra R, da Silvia SAG, Pinto SV, Dutra PML. Effect of exercise on the immune system: response, adaptation and cell signalling. *Review of Brazilian Journal of Sporting Medicine* 2012; 18(3):208–214.
23. Abbas AK, Lichtman AH. Cellular and molecular immunology. Philadelphia, PA: Elsevier Saunders 2015.
24. McCarthy DA, Dale MM. The leucocytosis of exercise. A review and model. *Sports Medicine* 1988; 6(6):333–363.
25. Fry RW, Morton AR, Crawford GP, Keast D. Cell numbers and in vitro responses of eucocytes and lymphocyte subpopulations following maximal exercise and interval training sessions of different intensities. *European Journal of Applied Physiology and Occupational Physiology* 1992; 64(3):218–227.
26. Muir AL, Cruz M, Martin BA, Thommasen H, Belzberg A, Hogg JC. Leukocyte kinetics in the human lung: role of exercise and catecholamines. *Journal of Applied Physiology* 1984; 57(3):711–719.
27. Van Eeden SF, Granton J, Hards JM, Moore B, Hogg JC. Expression of the cell adhesion molecules on leukocytes that demarginate during acute maximal exercise. *Journal of Applied Physiology* 1999; 86(3):970–976.
28. Nieman DC, Nehlsen-Caranella S. Effects of endurance exercise on the immune response. In Shepard RJ, Åstrand P-O (Eds). *Endurance in Sport*. London: Blackwell Science Ltd: 1992.p,487–504
29. Peake JM. Exercise-induced alterations in neutrophil degranulation and respiratory burst activity: possible mechanisms of action. *Exercise Immunology Review* 2002; 8:49–100.
30. Timmons BW, Cieslak T. Human Natural Killer Cell Subsets and Acute Exercise: A Brief Review. *Exercise Immunology Review* 2008; 8(14):8–23.
31. Nagao F, Suzui M, Takeda K, Yagita H, Okumura K. Mobilization of NK cells by exercise: down modulation of adhesion

- molecules on NK cells by catecholamines. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 2000; 279:R1251–1256.
32. Dela F, Mikines KJ, Von Linstow M, Galbo H. Heart rate and plasma catecholamines during 24 h of everyday life in trained and untrained men. *Journal Applied Physiology* 1992; 73:2389–2395.
  33. Jones B, Hoyne GF. The role of the innate and adaptive immunity in exercise induced muscle damage and repair. *Journal of Clinical and Cellular Immunology* 2017; 8(1):1000482.
  34. Gannon GA, Rhind SG, Suzui M, Shek PN, Shephard RJ. Circulating levels of peripheral blood leucocytes and cytokines following competitive cycling. *Cancer Journal of Applied Physiology* 1997; 22:133–147.
  35. Walsh N. Effect of oral glutamine supplementation on human neutrophil lipopolysaccharide-stimulated degranulation following prolonged exercise. *International Journal of Sport Nutrition and Exercise Metabolism* 2000; 10:39–50.
  36. Morozov VI., Tsyplenkov PV, Golberg ND, Kalinski MI. The effects of high-intensity exercise on skeletal muscle neutrophil myeloperoxidase in untrained and trained rats. *European Journal of Applied Physiology* 2006; 97:716–722.
  37. Woods JA, Lu Q, Ceddia MA, Lowder T. Exercise-induced modulation of macrophage function. *Immunology and Cell Biology* 2000; 78:545–553.
  38. Forner MA, Collazos ME, Barriga C, De La Fuente M, Rodriguez AB, Ortega E. Effect of age on adherence and chemotaxis capacities of peritoneal macrophages. Influence of physical activity stress. *Mechanisms of Ageing and Development* 1994; 75:179–189.
  39. Ortega E, Forner MA, Barriga C. Exercise-induced stimulation of murine macrophage chemotaxis: Role of corticosterone and prolactin as mediators. *Journal of Physiology (Lond.)* 1997; 498:729–734.
  40. Michna H. The human macrophage system: activity and functional morphology. *Bibliotheca Anatomica* 1988; 31:1–84.
  41. De La Fuente M, Martin MI, Ortega E. Changes in the phagocytic function of peritoneal macrophages from old mice after strenuous physical exercise. *Comparative Immunology. Microbiology and Infectious Diseases* 1990; 13:189–198.
  42. Ortega E, Rodriguez MJ, Barriga C, Forner MA. Corticosterone, prolactin and thyroid hormones as hormonal mediators of the stimulated phagocytic capacity of peritoneal macrophages after high-intensity exercise. *International Journal of Sports Medicine* 1996; 17:149–155.
  43. Woods JA, Davis JM, Mayer EP, Ghaffar A, Pate RR. Effects of exercise on macrophage activation for anti-tumor cytotoxicity. *Journal of Applied Physiology* 1994; 76:2177–2185.
  44. Weigent DA, Blalock JE. Associations between the neuroendocrine and immune systems. *Journal of Leukocyte Biology* 1995; 57:137–150.
  45. Kohut ML, Davis JM, Jackson DA, Colbert LH, Strasner A, Essig DA, Pate RR, Ghaffar A, Mayer EP. The role of stress hormones in exercise-induced suppression of alveolar macrophage anti-viral function. *Journal of Neuroimmunology* 1998; 81:193–200.
  46. Woods JA, Davis JM, Smith JA, Nieman DC. Exercise and cellular innate immune function. *Medicine and Science in Sports and Exercise* 1999; 31:57–66.
  47. Gabriel H, Urhausen A, Brechtel L, Muller HJ, Kindermann W. Alterations of regular and mature monocytes are distinct and dependent on intensity and duration of exercise. *European Journal of Applied Physiology* 1994; 69:179–181.
  48. Vollaard NB, Shearman JP, Cooper CE. Exercise-induced oxidative stress: myths, realities and physiological relevance. *Sports Medicine* 2005; 35:1045–1062.
  49. Barbieri E, Sestili P. Reactive oxygen species in skeletal muscle signaling. *Journal of Signal Transduction* 2012; 982794.
  50. Gomes EC, Silva AN, de Oliveira MR. Oxidants, antioxidants and the beneficial roles of exercise-induced production of reactive species. *Oxidative Medicine and Cellular Longevity* 2012; 756132.
  51. Nunes-Silva A, Freitas-Lima LC. The association between physical exercise and reactive oxygen species (ROS) production. *Sports Medicine and Doping Studies* 2015; 5(1):1000152.
  52. Babior BM. NADPH oxidase: an update. *Blood* 1999; 93:1464–1476.
  53. Tidball JG. Inflammatory processes in muscle injury and repair. *American Journal*



- of Physiology-Regulatory, Integrative and Comparative Physiology 2005; 288:R345–R353.
54. Peake JM., Suzuki K, Coombes JS. The influence of antioxidant supplementation on markers of inflammation and the relationship to oxidative stress after exercise. *Journal of Nutritional Biochemistry* 2007; 18:357–371.
  55. Dröge W. Free radicals in the physiological control of cell function. *Physiological Reviews* 2002; 82:47–95.
  56. Yang D, Elnor SG, Bian ZM, Till GO, Petty HR, Elnor VM. Pro-inflammatory cytokines increase reactive oxygen species through mitochondria and NADPH oxidase in cultured RPE cells. *Experimental Eye Research* 2007; 85(4):462–472.
  57. Duval C, Cantero AV, Auge N, Mabile L, Thiers JC, Negre-Salvayre A, Salvayre R. Proliferation and wound healing of vascular cells trigger the generation of extracellular reactive oxygen species and LDL oxidation. *Free Radical Biology and Medicine* 2003; 35:1589–1598.
  58. Kopprasch S, Pietzsch J, Graessler J. Validation of different chemiluminescent substrates for detecting extracellular generation of reactive oxygen species by phagocytes and endothelial cells. *Luminescence* 2003; 18:268–273.
  59. Jin CH, Paik IY, Kwak YS, Jee YS, Kim JY. Exhaustive submaximal endurance and resistance exercises induce temporary immunosuppression via physical and oxidative stress. *Journal of Exercise Rehabilitation* 2015; 11:198–203.
  60. Marzatico F, Pansarasa O, Bertorelli L, Somenzini L, DellaValle G. Blood free radical antioxidant enzymes and lipid peroxides following long distance and lactacidemic performances in highly trained aerobic and sprint athletes. *Journal of Sports Medicine and Physical Fitness* 1997; 37:235–239.
  61. Yavari A, Javadi M, Mirmiran P, Bahadoran Z. Exercise-induced oxidative stress and dietary antioxidants. *Asian Journal of Sports Medicine* 2015; 6:e24898.
  62. He F, Li J, Liu Z, Chuang CC, Yang W, Zuo L. Redox mechanism of reactive oxygen species in exercise. *Frontiers in Physiology* 2016; 7:486.
  63. Osburn WO, Kensler TW. Nrf2 signaling: an adaptive response pathway for protection against environmental toxic insults. *Mutation Research* 2008; 659:31–39.
  64. Muthusamy VR, Kannan S, Sadhaasivam K, Gounder SS, Davidson CJ, Boehme C, Hoidal JR, Wang L, Rajasekaran NS. Acute exercise stress activates Nrf2/ARE signalling and promotes antioxidant mechanisms in the myocardium. *Free Radical Biology and Medicine* 2012; 52: 366–376.
  65. Gleeson M, Pyne DB. Exercise effects on mucosal immunity. *Immunology and Cell Biology* 2000; 78:536–544.
  66. Simpson RJ, Kunz H, Agha N, Graff R. Molecular and cellular regulation of adaptation to exercise. In Claude Bouchard (Eds.), *Exercise and the Regulation of Immune Functions*. US. Elsevier: 2015. P.355–380.
  67. Moticka EJ. A Historical Perspective on Evidence-Based Immunology. In Edward J. Moticka (Eds.), *The Mucosal Immune System and Secretory IgA*. US. Elsevier: 2016. P.261–267.
  68. Brandtzaeg P, Baekkevold ES, Farstad IN, Jahnsen FL, Johansen FE, Nilsen EM, Yamanaka T. Regional specialisation in the mucosal immune system: What happens in the microcompartments? *Immunology Today* 1999; 20:141–51.
  69. Hanson LÅ, Bjökander J, Oxelius VA. Selective IgA deficiency. In: Chandra RK (ed.). *Primary and Secondary Immunodeficiency Disorders*. 1983.p.62–64 Edinburgh: Churchill Livingstone.
  70. Steerenberg PA, van Asperen IA, van Nieuw Amerongen A, Biewenga J, Mol D, Medema G. Salivary levels of immunoglobulin A in triathletes. *European Journal of Oral Science* 1997; 105:305–309.
  71. Bishop NC, Gleeson M. Acute and chronic effects of exercise on markers of mucosal immunity. *Frontier Bioscience (Landmark Ed.)* 2009; 14:4444–4456.
  72. Tharp GD. Basketball exercise and secretory immunoglobulinA. *European Journal of Applied Physiology* 1991; 63:312–314.
  73. Müns G, Singer P, Wolf F, Rubinstein I. Impaired nasal mucociliary clearance in long-distance runners. *International Journal of Sports Medicine* 1995; 16:209–213.
  74. Romagnani S. Type 1 T helper and type 2 T helper cells: functions, regulation and role in protection and disease. *International*

- Journal of Clinical Laboratory Research 1991; 21:152–158.
75. Gleeson M, Bishop NC, Stensel DJ, Lindley MR, Mastana SS, Nimmo MA. The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease. *Nature Reviews Immunology* 2011; 11:1–9.
  76. Vazquez MI, Catalan-Dibene J, Zlotnik A. B cells responses and cytokine production are regulated by their immune microenvironment. *Cytokine* 2015; 74(2):318–326.
  77. Gannon GA, Rhind SG, Suzui M, Shek PN, Shephard RJ. (1997). Circulating levels of peripheral blood leukocytes and cytokines following competitive cycling. *Canadian Journal of Applied Physiology* 1997; 22:133–147.
  78. Ostrowski K, Rohde T, Asp S, Schjerling P, Pedersen BK. Pro- and anti-inflammatory cytokine balance in strenuous exercise in humans. *Journal of Physiology (Lond.)* 1999; 515:287–291.
  79. Bruunsgaard H, Hartkopp A, Mohr T, Konradsen H, Heron I, Mordhorst CH, Pedersen BK. In vivo cell-mediated immunity and vaccination response following prolonged, intense exercise. *Medicine and Science in Sports and Exercise* 1997; 29:1176–1181.
  80. Pedersen BK., Febbraio MA. Muscle as an endocrine organ: focus on muscle-derived interleukin-6. *Physiological Reviews* 2008; 88:1379–1406.
  81. Pedersen BK. Muscles and their myokines. *The Journal of Experimental Biology* 214, 2011; 337–346.
  82. Janeway CA, Travers P, Walport M, Capra J. (2007); *Imunobiologia: O sistema imune na saúde e na doença – 6ed.* Editora Artmed.
  83. Yang Q, Guan KL. Expanding mTOR signaling. *Cell Research* 2007; 17(8):666–81.
  84. Thomson AW, Turnquist HR, Raimondi G. Immunoregulatory functions of mTOR inhibition. *Nature Immunology Review* 2009; 9(5):324–337.
  85. Wang J, Wolin MS, Hintze TH. Chronic exercise enhances endothelium-mediated dilation of epicardial coronary artery in conscious dogs. *Circulation Research* 1993; 73:829–838.
  86. Bowles DK, Wamhoff BR. Coronary smooth muscle adaptation to exercise: does it play a role in cardioprotection? *Acta Physiologica Scandinavica* 2003; 178:117–121.
  87. Shyy JY, Chien S. Role of integrins in endothelial mechanosensing of shear stress. *Cardiovascular Research* 2002; 91(9):769–775.
  88. Kojda G, Hambrecht R. Molecular mechanisms of vascular adaptations to exercise. Physical activity as an effective antioxidant therapy? *Cardiovascular Research* 2005; 67(2):187–197.
  89. Weichhart T, Costantino G, Poglitsch M, Rosner M, Zeyda M, Stuhlmeier KM., Kolbe T, Stulnig TM., Hörl WH, Hengstschläger M, Müller M, Säemann MD. The TSC–mTOR signaling pathway regulates the innate inflammatory response. *Immunology* 2008; 29:565–77.
  90. Handschin C, Spiegelman BM. The role of exercise and PGC1 $\alpha$  in inflammation and chronic disease. *Nature* 2008; 454:463–469.
  91. He C, Jr RS, Levine B. Exercise induces autophagy in peripheral tissues and in the brain. *Autophagy* 2012; 8(10):1548–1551.
  92. Levine B, Kroemer G. Autophagy in the pathogenesis of disease. *Cell* 2008; 132:27–42.
  93. Mizushima N, Levine B. Autophagy in mammalian development and differentiation. *Nature Cell Biology* 2010; 12:823–830.
  94. Chatterjee S, Sarkar S, Bhattacharya S. Toxic metals and autophagy. *Chemical Research in Toxicology* 2014; 27(11):1887–1900.
  95. Klionsky DJ, Cuervo AM, Seglen PO. Methods for monitoring autophagy from yeast to human. *Autophagy* 2007; 3:181–206.
  96. Mizushima N, Levine B, Cuervo AM, Klionsky DJ. Autophagy fights disease through cellular self digestion. *Nature* 2008; 451:1069–1074.
  97. Vainshtein A, Hood DA. The regulation of autophagy during exercise in skeletal muscle. *Journal of Applied Physiology* 2015; 120:664–673.
  98. Goodman CA, Mayhew DL, Hornberger TA. Recent progress toward understanding the molecular mechanisms that regulate skeletal muscle mass. *Cell Signaling* 2011; 23:1896–1906.
  99. Sanchez AM, Barbardi H, Py G, Candau RB. Autophagy is essential to support

- skeletal muscle plasticity in response to endurance exercise. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 2014; 307:R956–R969.
100. Ferraro E, Giammarioli AM, Chiandotto, S, Spoletini I, Rosano G. Exercise-induced skeletal muscle remodeling and metabolic adaptation: redox signaling and role of autophagy. *Antioxidants and Redox Signaling* 2014; 21:154–176.
101. Russ DW, Krause J, Wills A, Arreguin R. “SR stress” in mixed hind limb muscles of aging male rats. *Biogerontology* 2012; 13:547–555.
102. Mofarrahi M, Guo Y, Haspel JA, Choi AM, Davis EC, Gouspillou G, Hepple RT, Godin R, Burelle Y, Hussain SN. Autophagic flux and oxidative capacity of skeletal muscles during acute starvation. *Autophagy* 2013; 9:1604–1620.
103. Rahman M, Mofarrahi M, Kristof AS, Nkengfac B, Harel S, Hussain SNA. Reactive oxygen species regulation of autophagy in skeletal muscles. *Antioxidant and Redox Signaling* 2014; 20:443–459.
104. Hood DA, Ugucioni G, Vainshtein A, D’souza D. Mechanisms of exercise-induced mitochondrial biogenesis in skeletal muscle: implications for health and disease. *Comparative Physiology* 2011; 1:1119–1134.
105. Grumati P, Coletto L, Schiavinato A, Castagnaro S, Bertaggia E, Sandri M, Bonaldo P. Physical exercise stimulates autophagy in normal skeletal muscles but is detrimental for collagen VI-deficient muscles. *Autophagy* 2011; 7:1415–1423.
106. Wohlgenuth SE, Lees HA, Marzetti E, Manini TM, Aranda JM, Daniels MJ, Pahor M, Perri M. G, Leeuwenburgh C, Anton SD. An exploratory analysis of the effects of a weight loss plus exercise program on cellular quality control mechanisms in older overweight women. *Rejuvenation Research* 2011; 14:315–324.

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