

# Sputum as a Diagnostic Matrix for Respiratory Disease Screening

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## ABSTRACT

Mucus hyper-secretion is a common feature in most of respiratory diseases. Sputum contains white blood cells, cellular debris, dead tissue, serous fluid and offers an easy, inexpensive and non-invasive method of disease diagnosis. Sputum is rich in lipids, glycoconjugates and proteins. These molecules could be secreted from respiratory tract, lungs and infected sites as well as of pathogen origin. Complete sputum proteome profiling could be useful by adopting mass spectrometry methods to discover suitable clinical markers that could explain the disease and could be used for development of easy to use cost effective diagnostic test. The advancement of proteomics tools will be useful to carry out such biomarker discovery work. In this article, we will be discussing about the sputum proteome analysis in different respiratory disease conditions. Successful application of proteomics tool for understanding perturbed pulmonary disease conditions using sputum as a diagnostic matrix will be discussed.

**Key words:** Diagnosis, Noninvasive, Proteomics, Respiratory, Sputum.

## INTRODUCTION

Worldwide more than 1 billion people are reported to be affected by respiratory diseases every year. The main risk factors for respiratory diseases involving lower respiratory tract infection are use of tobacco, air pollution, occupational dust and chemical exposure. Most common respiratory diseases are chronic obstructive pulmonary disease (COPD), asthma, Cystic fibrosis, Lung Cancer, pulmonary hypertension and tuberculosis (TB).<sup>[1]</sup> Excessive mucus production in airways is a common feature of respiratory diseases. Expectoration is the act of coughing up or spitting out the material produced in the respiratory tract.<sup>[3]</sup>

When mucus is cough up from the lower airways is called sputum.<sup>[2]</sup> Primary role of mucus is to defend the respiratory tract from the noxious substance and to maintain airways from external damages. Secretion of mucus in airways is a homeostatic mechanism. It protects the respiratory tract from external particles and internal damages. Mucus is highly hydrated fluid layer that cover the mucosal surface. It is rich in secreted mucin and other molecules involved in host defense against infection. Mucin is secreted by goblet cells in epithelium and submucosal glands.<sup>[4]</sup> Mucin is 10-40 MDa in size and 3-10 nm in diameter, which is dense, with 25-30 carbohydrate chains per 100 amino acid

residues and constitute up to 80% of the dry weight. <sup>[5]</sup> Sputum is rich in mucin and commonly used for microbiological investigations for different respiratory disease conditions.

### **GENE INVOLVED IN SPUTUM PRODUCTION**

Almost twenty genes, found in chromosomes 1, 3, 4, 7, 11, 12 and 19 are reported to be responsible for mucin production in human. Nice (*MUC1, MUC2, MUC4, MUC5AC, MUC5B, MUC7, MUC8, MUC11, MUC13*) out these twenty genes are expressed in respiratory tract. These mucin genes are further grouped to membrane bound (*MUC1* and *MUC4*), secreted (*MUC2, MUC5AC, MUC5B, MUC7*) and rest three (*MUC3, MUC6, MUC8*) are yet to be categorized. <sup>[6-11]</sup>

### **MECHANISM OF SPUTUM PRODUCTION**

Over expression of mucin genes, excess production secondary to mucus cell hyperplasia, hypertrophy or even metaplasia or goblet cells or gland hyper secretion of formed and stored mucin in the airways might contribute in higher sputum production. Altered eicosanoids and lipid mediators in disease conditions, inflammatory mediators, environmental agents/pollutant, bacterial derived products, reactive oxygen and nitrogen species, ATP and UTP, cytokines might induce over expression of mucin genes. <sup>(12-20)</sup>

### **SPUTUM AS A DIAGNOSTIC MATRIX**

Due to its source, rich molecular details and primarily due to its noninvasive method of collection, sputum is used as the most common diagnostic matrix in many respiratory disease conditions. In fact, sputum color chart is used to find our presence of microorganisms like rusty color indicates presence of pneumococcal bacteria. Green color of sputum is caused by neutrophil myeloperoxidase, foamy white may come from obstruction or even edema,

frothy pink in pulmonary edema. <sup>[4]</sup> Sputum smear observed under microscope could help identification of many bacterial and fungal diseases. <sup>[4,6]</sup> Sputum culture provides the most sensitive and specific test results for multiple bacterial and fungal disease conditions like tuberculosis and pneumonia. Effect of therapeutic intervention may also be monitored by change in sputum color. Purulent containing pus, yellow-greenish (mucopurulent) color suggests that treatment with antibiotics can reduce symptoms. A white, milky or opaque (muroid) appearance often means that antibiotics may be ineffective in treating symptoms.

To find disease specific early biomarkers for important health conditions, before getting it complicated is important. Sputum as a diagnostic matrix has several advantages to other commonly used diagnostic matrix like serum or plasma due to their non-invasive method of collection. So a noninvasive method of diagnostic matrix collection with minimum anxiety, discomfort gains higher acceptance to patient to undergo health inspection. <sup>[21-23]</sup> Disease onset, progression and treatment outcome may be monitored by sputum biomarkers. <sup>[24,25]</sup> Sputum analysis can indicate the presence of microbes and the degree and type of inflammation in the air ways. <sup>[26]</sup> This makes sputum as a preferred and important matrix for diagnosis of various respiratory diseases.

### **SPUTUM PROTEOMICS APPROACH IN DISEASE DETECTION**

Sputum is rich in genetic materials of both host and pathogen origin. Analysis of genetic material by amplifying the signal through polymerase chain reaction pathogen specific primer is important. It involves requirement of cost intensive chemicals, instruments and needs specific training. These methods have low turnaround time to get results and minimize time for initiating treatment. Transcriptomics analysis provides interesting information on disease diagnosis and therapeutic outcomes. Like

analysis of Isocitrate lyase mRNA was found to be correlated well with bacterial load in sputum at time of diagnosis and during therapeutic interventions. Quantitative real time polymerase chain reaction is useful for monitoring important analytes (mRNA) however it is cost intensive and requires high end instruments at temperature-controlled environment. Protein profiling has a potential advantage of discovering suitable clinical biomarkers that are significantly deregulated by disease and receiving treatment. [27] Identification of important proteins provide useful translational potential to develop easily deployable solutions like lateral flow devices that could be used in point of care for disease screening or treatment monitoring. Employing advanced quantitative proteomics tools, deregulated proteins expressed in diverse biofluids like plasma, serum, urine, sputum or saliva in disease conditions could be identified as important marker molecules. These marker molecules, after careful validation in independent test populations of diverse ethnic background could be useful to develop clinical assay for monitoring pathogenesis for respiratory diseases Fig. 1. [28,29]

### Tuberculosis

Majority of the pulmonary tuberculosis diagnosis is done using a 120 years old sputum acid fast microscopy test. Sputum microscopy test has significantly low sensitivity and specificity but due to its easy availability in resource limited conditions it is the important diagnosis test. Growing the tuberculosis causing microorganisms in sputum provides the highest sensitivity and specificity but suffer limited use due to long turnaround time and resource intensive procedure. A protein-based tuberculosis diagnostic method might be useful to develop an easy to use TB diagnostic and monitoring test to determine the treatment efficacy. [30-32] Sputum proteins from active tuberculosis and control subjects, separated in a two-

dimensional electrophoresis unit identified 62 differentially expressed protein spots. These proteins spots were analyzed using matrix-assisted laser desorption ionization time-of-flight/time-of-flight mass spectrometry (MALDI-TOF/TOF) and found to be 47 proteins were down regulated and 15 proteins were up-regulated. Most of these deregulated proteins were involved in acute phase response, signal transduction, cytoskeleton structure and immune response. Increase in abundance of acute phase proteins is to improve survival and restore haemostatic in host system. Among those deregulated proteins, eleven were signal-related proteins, six cytoskeleton-related proteins, five immune response-related proteins, four bacteriostatic proteins, 11 hypothetical human proteins and interestingly, a set of 7 bacterial proteins were identified in the sputum of active TB patients. Out of those 7 bacterial proteins MT3876 is a hypothetical MTB protein, which function is still unknown. Further validation of this MT3876 may be use as a biomarker for tuberculosis. [33]

### COPD

COPD is a common lung disease affects 8%-10% of adult of the developing World. [34] COPD is a polygenic disease and characterized by the chronic air flow obstruction and a range of pathological changes in the lung, some significant extra pulmonary effects such as cardiovascular, mental, systematic inflammation, anemia and musculoskeletal comorbidities may be contributed to the severities of the disease in individual patients. [35] The disease prevalence, morbidity, and mortality vary across countries and across different groups within countries. COPD is also influenced by other risk factors like deficiency of alpha-1 antitrypsin4, a major circulating inhibitor of serine proteases and other direct effect of tobacco smoking, burning of wood and other biomass fuels. [2,36] Sputum proteome of normal (healthy smokers) to chronic bronchitis, chronic obstructive pulmonary disease (COPD), and COPD

with emphysema were analyzed using CapLC-ESI-Q/TOF-MS identified a total of 203 host proteins. These proteins also showed evidence of disease progression from healthy to more advanced stage.

Zinc- $\alpha$ -2-glycoprotein,  $\beta$ -microsemino protein, cystatin S, and transthyretin shows significant variation in Smokers and Mild-to-Moderate COPD. [37] In a separate study, sputum samples processed in a two-dimensional poly acrylamide gel chromatography, 1325 individual spots were identified in COPD and control subjects. Out of which 37 were quantitatively and 3 were qualitatively showed significant difference between these study groups. Fifteen proteins were identified from the 40 protein spots analyzed using tandem mass spectrometry analysis. Seven of these important proteins were further quantified in induced sputum from 97 study individuals. Using this sequential approach, two potential biomarkers i.e., apolipoprotein A1 and

lipocalin-1 were reported to be significantly reduced in patients with COPD when compared with healthy smokers. Abundance of these important proteins was correlated with FEV<sub>1</sub>/FVC, indicating their relationship to disease severity. [38] In a recent study, 13 significantly deregulated proteins were identified in COPD patients using TandemMass Tag™6-plex (TMTsixplex™) and liquid chromatography tandem mass-spectrometry (LC-MS/MS) using an EASY-nanoLC 1000 instrument connected online to a QExactive™ mass-analyzer (Thermo Scientific) . Among which Keratin, type I cytoskeletal 19, UPF0762 protein C6orf58, Metalloproteinase inhibitor 1, BPI fold-containing family B member 1, Peptidyl-prolyl cis-trans isomerase B are upregulated and Serotransferrin, Alpha-2-HS-glycoprotein, Antithrombin-III, Afamin, Serum albumin, Histidine-rich glycoprotein, Apolipoprotein A-I, Beta-Ala-His dipeptidase are down regulated. [39]

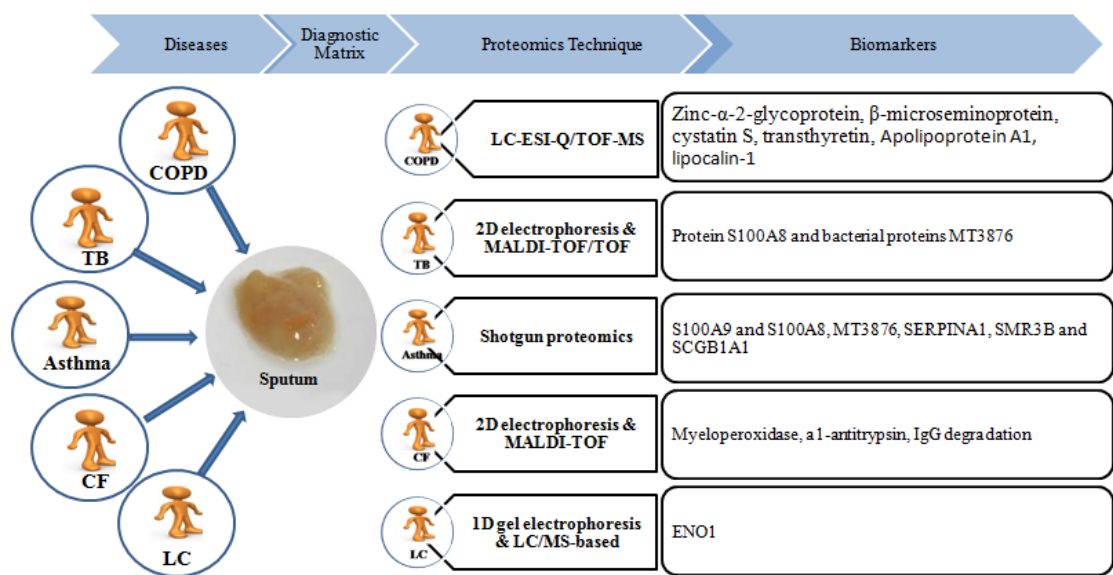


FIG. 1: Identified Proteomic Biomarkers In Different Respiratory Diseases Conditions Using Sputum As Matrix.

### Lung cancer

An estimated 1.8 million new lung cancer cases are reported every year accounting for 13% of all form of cancer diagnosis worldwide. It is the most common cause of cancer deaths. Lung cancer symptoms manifests usually very late or in

the advanced stage with symptoms of persistence cough, sputum with blood, chest pain, voice change, shortness of breath and recurrent pneumonia or bronchitis. [40] The prognosis for the patients with lung cancer is strongly correlated to the stage of the disease at the time of diagnosis. Whereas

patients with clinical stage IA, disease have a 5-year survival of about 60%, the clinical stage II-IV disease 5-year survival rate ranges from 40% to < 5%. [41] Over two-thirds of the patients have regional lymph-node involvement or distant disease at the time of presentation. [42] In sputum of lung cancer patients, majority of the protein compositions were from cancer sites.

Approximately 85% of lung tumors are non-small cell lung cancers (NSCLCs), and it has two major histological subtypes: squamous cell carcinoma (SCC) and adenocarcinoma (AC). [43] Mortality reduction of Lung cancer depends on the development of noninvasive approaches for early detection of NSCLC followed by suitable treatments. Cytological and molecular studies of exfoliated bronchial epitheliums in sputum were more accurate to identify SCC tumors that largely located in central areas of the lungs. [44-46] Cytological techniques have low sensitivity and specificity for the diagnosis of ACs, which are the most common type in peripheral region of the lungs. However, a cell surface protein ENO1 was identified as a marker by 1D gel electrophoresis and LC/MS-based proteomics in sputum sample of lung cancer patients. Later it was validated using western blotting and ELISA. The upregulated ENO1 in sputum sample has similar sensitivity and specificity for diagnosis of both SCCs and ACs. So, ENO1 could be used as a potential biomarker for detection of SCCs and ACs. [47]

### **Cystic Fibrosis**

Cystic fibrosis (CF) is a fatal genetic disease with incident of 1 in every 2,500. CF is caused by mutation in gene encoding CF transmembrane conductance regulator (CFTR) with symptoms of persistence coughing with phlegm, wheezing, and shortness of breath. In CF due to chronic bacterial infection, frequent exacerbation the patient die of respiratory failure. [48] Bacterial colonization in airways results submucosal hypertrophy and excessive mucus secretion in lung. [49] CF progression

over time is complex and is associated with infectious colonization of lungs, nutritional, environmental and social variables. Therapy depends on the symptoms, lung function and radiological changes which are lag behind the occurrence of established lung pathology. Induce sputum protein profiling from adult CF patients in comparison to healthy human and children with CF were studied using 2DE-PAGE and the protein spots with differential intensity were identified using MALDI-TOF. A set of protein panel including myeloperoxidase,  $\alpha$ 1-antitrypsin, IgG degradation, and total protein concentration in comparison with IL-8 showed differential expression which can be used as biomarker in lung exacerbation for diagnosis and prognosis of CF. [50]

### **Asthma**

In 2017, around 334 million people reported with symptoms of asthma around the world. Combination of genetic and environmental factors is mainly responsible for Asthma. Its diagnosis is usually based on the pattern of symptoms, response to therapy over time, and spirometry. Asthma is classified on the basis of the frequency of symptoms, forced expiratory volume in one second (FEV1), and peak expiratory flow rate. In asthma sputum hypersecretion contributes to the development of viscid mucus plugs that can occlude even the large airways and in chronic bronchitis it contributes to early morning coughing, sputum production and airways obstruction. In shotgun proteomics study 240 important proteins were identified from induce sputum of asthmatic patients. Most of these proteins were involved in defense response, protease inhibitor activity, immunity, response to inflammation and complement activation. Out of these 240 identified proteins, seventeen were differentially expressed between healthy and asthmatic group and involved in defense and stress response proteins from extracellular space. Among them calcium binding proteins S100A9 and S100A8, SERPINA1, SMR3B and

SCGB1A1. [51] Mouthuy et al. showed IgE levels are not related to disease severity but clearly increased in those exhibiting airway eosinophilic inflammation. The role of IgE has been traditionally assigned to allergic reaction towards an aeroallergen in sensitized patients. Humbert et al have drawn attention to the potential role of IgE in non-atopic asthma by showing increased expression of the receptor FcεRI in the bronchial mucosa in asthmatics irrespective of the atopic status. [52] Here they found that GRO-α, eotaxin-2, and Pulmonary and activation-regulated chemokine (PARC) were increased significantly in sputum specimens from patients with asthma. In particular, PARC in the airways may play an important role in the eosinophilic inflammation of asthmatic airways. PARC is elevated in sputum specimens from patients with asthma and may play important roles in development of airway eosinophilic inflammation in asthma. [53]

## CONCLUSION

Sputum as a diagnostic matrix for several respiratory disease conditions have showed high potential and may also be useful to monitor therapeutic outcome. Due to noninvasive mode of sample collection, it also possesses enormous translational value for clinical application. In this chapter we discussed the importance of different sputum proteins as marker for diagnosis of different respiratory disease conditions. The significance of recent advancements in the potential application of proteomic profile analyses of sputum has been highlighted in many common respiratory disease conditions. Sputum proteomics have been a novel approach in search for protein biomarker from different biofluids including sputum to detect human diseases. Comprehensive analysis and identification of the proteome content of sputum may also contribute to the understanding of the pathophysiology for respiratory disease conditions.

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### Competing interests

Authors have declared that no competing interests exist.

### Authors' Contributions

SCB, RKN and RKB wrote the review.

### Consent (where ever applicable)

Not applicable

### Ethical approval (where ever applicable)

Not applicable

## REFERENCES

1. Bousquet J, Dahl R, Khaltsev N. Global alliance against chronic respiratory diseases. *Allergy*. 2007; 62(3):216-23.
2. Abbey DE, Burchette RJ, Knutsen SF, McDonnell WF, Lebowitz MD, Enright PL. Long-term particulate and other air pollutants and lung function in nonsmokers. *Am J Respir Crit Care Med*. 1998; 158(1):289-98.
3. Rose MC, Voynow JA. Respiratory tract mucin genes and mucin glycoproteins in health and disease. *Physiol Rev*. 2006; 86(1):245-78.
4. Rose MC, Nickola TJ, Voynow JA. Airway mucus obstruction: mucin glycoproteins, MUC gene regulation and goblet cell hyperplasia. *Am J Respir Cell Mol Biol*. 2001; 25(5):533-7.
5. Lamblin G, Lhermitte M, Klein A, Houdret N, Scharfman A, Ramphal R, et al. The carbohydrate diversity of human respiratory mucins: a protection of the underlying mucosa. *Am Rev Respir Dis*. 1991; 144(3 Pt 2):S19-S24.
6. Rogers DF. Physiology of airway mucus secretion and pathophysiology of hypersecretion. *Respir Care*. 2007; 52(9):1134-49.
7. Gourevitch M, Von Mensdorff-Pouilly S, Litvinov SV, Kenemans P, van Kamp GJ, Verstraeten AA, Hilgers A. Polymorphic epithelial mucin (MUC-1)-containing circulating immune complexes in carcinoma patients. *Br J Cancer*. 1995; 72(4):934-38.
8. Li D, Gallup M, Fan N, Szymkowski DE, Basbaum CB. Cloning of the amino-terminal and 5'-Flanking region of the human MUC5AC mucin gene and transcriptional up-regulation by bacterial exoproducts. *J Biol Chem*. 1998; 273(12):6812-20.
9. Moniaux N, Nollet S, Porchet N, Degand P, Laine A, Aubert J-P. Complete sequence of the human mucin MUC4: a putative cell

- membrane-associated mucin. *Biochem J.* 1999; 338(2):325-33.
10. Shankar V, Pichan P, Eddy RL, Tonk V, Nowak NJ, Sait SNJ, et al. Chromosomal localization of a human mucin gene (MUC8) and cloning of the cDNA corresponding to the carboxy terminus. *Am J Respir Cell Mol Biol.* 1997; 16(3):232-41.
  11. Gum JR, Ho JJJ, Pratt WS, Hicks JW, Hill AS, Vinall LE, Robertson AM, Swallow DM, Kim YS. MUC3 human intestinal mucin analysis of gene structure, the carboxyl terminus, and a novel upstream repetitive region. *J Biol Chem.* 1997; 272(42):26678-86.
  12. Pigny P, Guyonnet-Duperat V, Hill AS, Pratt WS, Galiegue-Zouitina S, d'Hooge MC, et al. Human mucin genes assigned to 11p15. 5: identification and organization of a cluster of genes. *Genomics.* 1996; 38(3):340-52.
  13. Kim KC, Wasano K, Niles RM, Schuster JE, Stone PJ, Brody JS. Human neutrophil elastase releases cell surface mucins from primary cultures of hamster tracheal epithelial cells. *Proceedings of the National Academy of Sciences.* 1987; 84(24):9304-8.
  14. Samet JM, Cheng P-W. The role of airway mucus in pulmonary toxicology. *Environ Health Perspect.* 1994; 102(2):89-103.
  15. Longphre M, Li D, Li J, Matovinovic E, Gallup M, Samet JM, Basbaum CB. Lung mucin production is stimulated by the air pollutant residual oil fly ash. *Toxicol Appl Pharmacol.* 2000; 162(2):86-92.
  16. Adler KB, Hendley DD, Davis GS. Bacteria associated with obstructive pulmonary disease elaborate extracellular products that stimulate mucin secretion by explants of guinea pig airways. *Am J Pathol.* 1986; 125(3):501-14.
  17. Adler KB, Holden-Stauffer WJ, Repine JE. Oxygen metabolites stimulate release of high-molecular-weight glycoconjugates by cell and organ cultures of rodent respiratory epithelium via an arachidonic acid-dependent mechanism. *J Clin Invest.* 1990; 85(1):75-85.
  18. Fischer BM, Krunkosky TM, Wright DT, Dolan-O'Keefe M, Adler KB. Tumor necrosis factor- $\alpha$  stimulates mucin secretion and gene expression in airway epithelium in vitro. *Chest.* 1995;107(3):133S-5S.
  19. Abdullah L, Conway J, Cohn J, Davis C. Protein kinase C and Ca<sup>2+</sup> activation of mucin secretion in airway goblet cells. *Am J Physiol-Lung Cell Mol Physiol.* 1997; 273(1):L201-L10.
  20. Kim K, Park H, Shin C, Akiyama T, Ko K. Nucleotide-induced mucin release from primary hamster tracheal surface epithelial cells involves the P2u purinoceptor. *Eur Respir J.* 1996; 9(3):542-8.
  21. Zhang A, Sun H, Han Y, Yuan Y, Wang P, Song G, et al. Exploratory urinary metabolic biomarkers and pathways using UPLC-Q-TOF-HDMS coupled with pattern recognition approach. *Analyst.* 2012;137(18):4200-8.
  22. Wang X, Zhang A, Han Y, Wang P, Sun H, Song G, et al. Urine metabolomics analysis for biomarker discovery and detection of jaundice syndrome in patients with liver disease. *Mol Cell Proteomics.* 2012; 11(8):370-80.
  23. Kumada Y, Zhao C, Ishimura R, Imanaka H, Imamura K, Nakanishi K. Protein-protein interaction analysis using an affinity peptide tag and hydrophilic polystyrene plate. *Journal of Biotechnol.* 2007; 128(2):354-61.
  24. Wang W, Sun J, Hollmann R, Zeng A-P, Deckwer W-D. Proteomic characterization of transient expression and secretion of a stress-related metalloprotease in high cell density culture of *Bacillus megaterium*. *J Biotechnol.* 2006; 126(3):313-24.
  25. Forsberg L, Larsson C, Sofiadis A, Lewensohn R, Höög A, Lehtiö J. Prefractionation of archival frozen tumours for proteomics applications. *J Biotechnol.* 2006; 126(4):582-6.
  26. Lacy P, Lee JL, Vethanayagam D. Sputum analysis in diagnosis and management of obstructive airway diseases. *Ther Clin Risk Manag.* 2005; 1(3):169-79.
  27. Zhang A, Sun H, Wang P, Wang X. Salivary proteomics in biomedical research. *Clin Chim Acta.* 2013; 415:261-5.
  28. Lau AT, Chiu JF. Biomarkers of lung-related diseases: Current knowledge by proteomic approaches. *J Cell Physiol.* 2009; 221(3):535-43.
  29. Hirsch J, Hansen KC, Burlingame AL, Matthay MA. Proteomics: current techniques and potential applications to lung disease. *Am J Physiol-Lung Cell Mol Physiol.* 2004; 287(1):L1-L23.
  30. Wallis RS, Pai M, Menzies D, Doherty TM, Walzl G, Perkins MD, et al. Biomarkers and diagnostics for tuberculosis: progress, needs, and translation into practice. *Lancet.* 2010; 375(9729):1920-37.
  31. Nahid P, Kim PS, Evans CA, Alland D, Barer M, Diefenbach J, et al. Clinical research and development of tuberculosis diagnostics: moving from silos to synergy. *Journal of Infectious Diseases.* 2012; 205(2):S159-S68.
  32. McNerney R, Daley P. Towards a point-of-care test for active tuberculosis: obstacles and

- opportunities. *Nat Rev Microbiol.* 2011; 9(3):204-13.
33. Fu Y, Yi Z, Guan S, Zhang S, Li M. Proteomic analysis of sputum in patients with active pulmonary tuberculosis. *Clin Microbiol Infect.* 2012; 18(12):1241-7.
  34. Diaz-Guzman E, Mannino DM. Epidemiology and prevalence of chronic obstructive pulmonary disease. *Clin Chest Med.* 2014; 35(1):7-16.
  35. Watz H, Waschki B, Boehme C, Claussen M, Meyer T, Magnussen H. Extrapulmonary effects of chronic obstructive pulmonary disease on physical activity: a cross-sectional study. *Am J Respir Crit Care Med.* 2008; 177(7):743-51.
  36. Jindal S, Aggarwal A, Chaudhry K, Chhabra S, D Souza G, Gupta D, et al. A multicentric study on epidemiology of chronic obstructive pulmonary disease and its relationship with tobacco smoking and environmental tobacco smoke exposure. *Indian J Chest Dis Allied Sci.* 2006; 48(1):23-29.
  37. Casado B, Iadarola P, Pannell LK, Luisetti M, Corsico A, Ansaldo E, et al. Protein expression in sputum of smokers and chronic obstructive pulmonary disease patients: a pilot study by CapLC-ESI-Q-TOF. *J Proteome Res.* 2007; 6(12):4615-23.
  38. Nicholas B, Skipp P, Mould R, Rennard S, Davies DE, O'Connor CD, et al. Shotgun proteomic analysis of human-induced sputum. *Proteomics.* 2006; 6(15):4390-401.
  39. Titz B, Sewer A, Schneider T, Elamin A, Martin F, Dijon S, et al. Alterations in the sputum proteome and transcriptome in smokers and early-stage COPD subjects. *J Proteomics.* 2015; 128:306-20.
  40. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. *CA Cancer J Clin.* 2015; 65(1):5-29.
  41. Mountain CF. Revisions in the international system for staging lung cancer. *Chest.* 1997; 111(6): 1710-7.
  42. Ihde DC. Chemotherapy of lung cancer. *New England Journal of Medicine.* 1992; 327(20): 1434-41.
  43. Minna JD, Roth JA, Gazdar AF. Focus on Lung Cancer. *Cancer Cell.* 2002; 1(1):49-52.
  44. Varella-Garcia M, Schulte AP, Wolf HJ, Feser WJ, Zeng C, Braudrick S, et al. The detection of chromosomal aneusomy by fluorescence in situ hybridization in sputum predicts lung cancer incidence. *Cancer Prev Res.* 2010; 3(4):447-53
  45. Li R, Todd NW, Qiu Q, Fan T, Zhao RY, Rodgers WH, et al. Genetic Deletions in Sputum as Diagnostic Markers for Early Detection of Stage I Non-Small Cell Lung Cancer. *Clin Cancer Res.* 2007; 13(2):482-7.
  46. Fjiang Jiang F, Todd N, Li R, Zhang H, Fang H-B, Stass SA. A panel of sputum-based genomic marker for early detection of lung cancer. *Cancer Prev Res.* 2010; 3(12):1571-8.
  47. Yu L, Shen J, Mannoor K, Guarnera M, Jiang F. Identification of ENO1 as a potential sputum biomarker for early-stage lung cancer by shotgun proteomics. *Clin Lung Cancer.* 2014; 15(5):372-8. e1.
  48. Elborn JS. Cystic fibrosis. *Lancet.* 2016; 388(10059):2519-31.
  49. Gibson RL, Burns JL, Ramsey BW. Pathophysiology and management of pulmonary infections in cystic fibrosis. *Am J Respir Crit Care Med.* 2003; 168(8):918-51.
  50. Sloane AJ, Lindner RA, Prasad SS, Sebastian LT, Pedersen SK, Robinson M, et al. Proteomic analysis of sputum from adults and children with cystic fibrosis and from control subjects. *Am J Respir Crit Care Med.* 2005; 172(11):1416-26.
  51. Gharib SA, Nguyen EV, Lai Y, Plampin JD, Goodlett DR, Hallstrand TS. Induced sputum proteome in healthy subjects and asthmatic patients. *J Allergy Clin Immunol.* 2011; 128(6):1176-84. e6.
  52. Mouthuy J, Detry B, Sohy C, Pirson F, Pilette C. Presence in Sputum of Functional Dust Mite-Specific IgE Antibodies in Intrinsic Asthma. *Am J Respir Crit Care Med.* 2011;184(2):206-14.
  53. Kim H-B, Kim C-K, Iijima K, Kobayashi T, Kita H. Protein microarray analysis in patients with asthma: elevation of the chemokine PARC/CCL18 in sputum. *Chest.* 2009; 135(2):295-302.

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