

# Caveolae and Caveolin: Potential Targets for Cardioprotection

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## ABSTRACT

Caveolae are sphingolipid and cholesterol rich micro-domains of the plasma membrane that coordinate and regulate varieties of signaling processes. Caveolae present in essentially all cell types of the cardiovascular system, including endothelial cells, smooth muscle cells, macrophages, cardiac myocytes, and fibroblasts. Numerous functions have been ascribed to this omega-shaped sphingolipid and cholesterol rich micro-domains.

Caveolae are receiving increasing attention as cellular organelles contributing to the pathogenesis of several structural and functional processes including cardiac hypertrophy and heart failure. At present, very little is known about the role of caveolae in cardiac function and diseases, although recent studies with caveolin knock-out mouse have shown that caveolae and caveolins play a pivotal role in various human pathobiological conditions. This review will discuss the possible role of caveolae in cardiac health and disease.

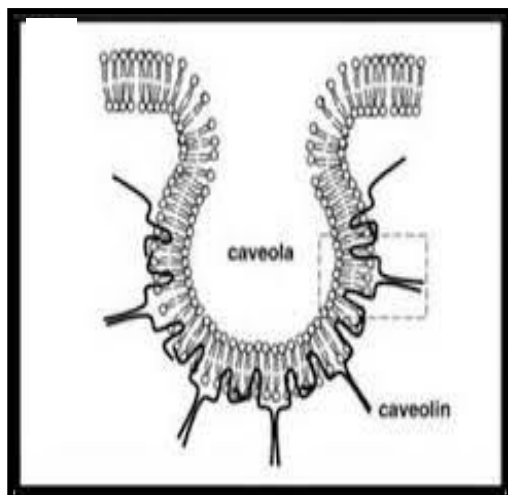
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## INTRODUCTION

The “fluid-mosaic model” suggested by Singer and Nicolson in 1972 described the membrane as a fluid bilayer with a homogeneous lipid distribution. In recent years this model has been refined as the plasma membrane, which has been shown to consist of defined lipid rich regions interspersed with more fluidic membrane regions resulting in a much more complex view of the membrane than originally hypothesized. Caveolae are subcellular structures that were first described using electron microscopy in 1953 by George Palade. Initially, these invaginations were identified as plasmalemmal vesicles. Two years later in 1955, similar structures were reported by Yamada E, in the gall bladder epithelium, and were described as caveolae intracellulares due to their cave-like, invaginated appearance [Fig-1]. Since their initial discovery, caveolae have been found

in almost all cell types. Recent studies have shown that caveolar microdomains are more than lipid enriched invaginations of the plasma membrane. <sup>[1]</sup> Caveolae play an important role in physiological functions such as cell signaling, <sup>[2,3]</sup> endocytosis, calcium homeostasis, adrenergic receptor regulation, and intracellular cholesterol transport. <sup>[4-6]</sup>

Caveolins, structural proteins essential for caveolae formation, are present in three isoforms (Cav-1, Cav-2 and Cav-3). Cav-1 was the first member of the caveolin family to be identified as a phosphorylated protein. Caveolins are a family of 21- to 25-kDa integral membrane proteins that have been implicated in a variety of cellular functions. <sup>[1]</sup> Currently, three caveolin genes are known to exist. Caveolin-1 and caveolin-2 are ubiquitously expressed, while the expression of caveolin-3 is muscle specific. <sup>[7,8]</sup>



**Fig 1- Diagrammatic representation of caveolae.**  
 [from : P. L. Tuma, A. L. Hubbard. (2003) Transcytosis: Crossing cellular barriers. *Physiological Reviews*. 83 (3):871-932]

### Caveolae and Cellular Signaling Components

A large number of GPCR (G-protein coupled receptor) have been reported to co-localize with caveolae. In case of Angiotensin I receptor, GPCR-caveolin interaction is important for receptor sorting and delivery to plasma membrane. [9] According to the caveolin signaling hypothesis, caveolae bring downstream effectors in proximity to receptors (e.g., GPCRs) for initiating receptor, tissue and cell-specific signal transduction. [10,11] These effectors are thought to reside within caveolae by direct interaction with caveolin. Palmitoylation may enhance caveolar localization of proteins. [12,13]

Among the different binding proteins of caveolin, its interaction with eNOS has been most extensively studied. [14] Binding of eNOS with caveolin inhibits enzyme activity [15] and loss of caveolin expression upregulates eNOS activity. [16] Like eNOS, caveolin is also thought to negatively regulate Adenylate Cyclase (AC) activity. Caveolin-1 and caveolin-3, but not caveolin-2 inhibits AC activity and this inhibition is AC isoforms. [17] Like eNOS, protein kinases (PKA/PKC) can also interact with caveolin-1 and inhibit its activity. [18] The PKC family of enzymes translocate to the cellular compartment in response to the external stimuli. [19] The

phosphatidylinositol-3-kinase/protein kinase B (PI3K/PKB, Akt) pathway is another protein kinase system that interacts with caveolin and this interaction may regulate cell survival. For example, caveolin retains Akt in activated form (phosphorylated form) in prostate cancer, [20] presumably via interaction with caveolin scaffolding domain of caveolin and by inhibition of protein phosphatase 1 and 2A. [21] In muscle, we can also found a linear relationship between the expression of caveolin-3 and activation of PI3K/Akt pathway in the regulation of cell survival. [22] In addition, the phosphorylated form of caveolin is involved in EGF receptor transactivation, which is dependent on Src and Akt phosphorylation and for which caveolin helps integrate this signaling cascade. [23]

Receptor tyrosine kinases also have been localized to caveolae [e.g., EGF, NGF, IGF and PGDF] and their downstream effectors MAP kinases, which regulate numerous cellular processes, are also regulated by caveolin. [24,25] P42/44 MAPK localizes to caveolae and is negatively regulated by interaction with caveolin 1. [26] Overexpression of caveolin-1 also inhibits the MEK/ERK signaling pathways. [27] Consistent with this action, caveolin-1 and-3 knockout mice showed increased activation of p42/44 MAPK. [28] Ischemia reperfusion showed differential activation of p42/44 ERK and p38MAPK in caveolar and noncaveolar fraction, indicating differential regulation of these kinases by caveolin. [29] Certain non-receptor tyrosine kinases such as members of src family (c-Src, Fyn, lyn) are enriched in caveolae and interactions with caveolin-1 also suppress the kinases activities. [30,31] Tyrosine phosphorylation of caveolin itself makes phospho caveolin, which acts as a key site of tyrosine kinase signaling. [32]

### Caveolin Knockout and Different Cardiomyopathies

The elucidation of the role of caveolae has been the topic of many investigations which were greatly enhanced

after the discovery of caveolin, the protein marker of these flask-shaped plasma membrane invaginations. The generation of mice deficient in the various caveolin genes (cav-1, cav-2 and cav-3) has provided physiological models to unravel the role of caveolins or caveolae at the whole organism level.

Caveolin-KO mice (Cav-1,-2, -3) and caveolin 1/3 double KO mice have already been developed. Although they are viable, they are fertile but display numerous phenotypes. Caveolin-1 knockout mice develop progressive cardiac hypertrophy as demonstrated by transthoracic echocardiography (TTE) and magnetic resonance imaging (MRI).<sup>[28]</sup> In contrast, caveolin-3 knockout mice develop cardiomyopathy characterized by hypertrophy, vasodilatation and reduced contractility as well.<sup>[33]</sup> Caveolin-1 and caveolin-3 double knockout mice completely lacking caveolae are deficient in all three caveolin proteins because caveolin-2 is degraded in absence of caveolin-1. The double knockout mice developed severe cardiomyopathic phenotype with cardiac hypertrophy and decreased contractility.<sup>[34]</sup> Additionally, Cav-1 KO mice exhibited myocardial hypertrophy, pulmonary hypertension and alveolar cell hyper proliferation caused by constitutive activation of p42/44 mitogen activated protein kinase and Akt.<sup>[35]</sup> Interestingly, in Cav-1-reconstituted mice, cardiac hypertrophy and pulmonary hypertension were completely rescued.<sup>[35]</sup> Again, genetic ablation of Cav-1 leads to a striking biventricular hypertrophy and to a sustained eNOS hyper-activation yielding increased systemic NO levels.<sup>[36]</sup> Furthermore, a diminished ATP content and reduced level of cyclic AMP in hearts of knockout mice was also reported.<sup>[36]</sup> Taken together, these results indicate that genetic disruption of caveolin-1 is sufficient to induce severe biventricular hypertrophy with signs of systolic and diastolic heart failure.<sup>[36]</sup>

Apart from its ability to degrade extracellular matrix proteins, matrix

metalloproteinase-2 (MMP-2) was recently revealed to have targets and actions within the cardiac myocyte. MMP-2 (gelatinase A) has been localized to the thin and thick myofilaments of the cardiac sarcomere, as well as to the nucleus.<sup>[37,38]</sup> The intracellular proteins troponin I and myosin light chain-1 are proteolyzed by MMP-2 in ischemia/reperfusion injury.<sup>[37,38]</sup> The tissue inhibitors of metalloproteinase (TIMPs) control MMP activities,<sup>[39]</sup> but other mechanisms of regulation are less well elucidated. In endothelial cells, MMP-2 has been localized to the caveolae<sup>[40]</sup> yet its function there is unknown. Disruption of caveolae activates MMP-2 in fibrosarcoma cells<sup>[41]</sup> while Cav-1 overexpression in tumor cells causes decreased MMP-2 activity<sup>[42]</sup> suggesting that Cav-1 may participate in the regulation of MMP-2. Whether the role of MMP-2 activity in the heart is affected by caveolin still remains unknown. Here we present evidence that MMP-2 localizes with Cav-1 in the mouse heart, and that CSD inhibits MMP-2 activity and that hearts of mice deficient in Cav-1 have increased MMP-2 activity.

Interestingly, Cav-3 KO mice show a number of myopathic changes, consistent with a mild to moderate muscular dystrophy phenotype. However, it remains unknown whether a loss of cav-3 affects the phenotypic behavior of cardiac myocytes in vivo. Cav-3 knockout hearts display significant hypertrophy, dilation and reduced fractional shortening as revealed by gated cardiac MRI and transthoracic echocardiography. Histological analysis reveals marked cardiac myocyte hypertrophy, with accompanying cellular infiltrates and progressive interstitial/ perivascular fibrosis. It has also demonstrated that p42/44MAPK (ERK1/2) is hyperactivated in heart derived from caveolin-3 knockout mice, which can lead to cardiac hypertrophy.<sup>[43]</sup>

In the endoplasmic reticulum, Cav-3 initiates the biogenesis of caveolae organelles by forming homo-oligomers and hetero-oligomers with Cav-1.<sup>[44]</sup> At the

plasmalemma, Cav-3 interacts with dystrophin and its associated glycoproteins. [45,46] Cav-3 and dystrophin competitively bind to the same site of  $\beta$ -dystroglycan, suggesting that Cav-3 may regulate the membrane recruitment of dystrophin and the assembly of the dystrophin glycoprotein complex (DGC). [47] At the cell surface, Cav-3 colocalizes also with signaling molecules such as  $G_i2\alpha$ ,  $G_{\beta\gamma}$ , c-Src, other Src kinases as well as nitric oxide synthases (neuronal and inducible NOS), indicating that muscle caveolae might be involved in the modulation of these signaling processes. [48,49] In addition, Cav-3 plays a role in the regulation of energy metabolism of muscle cells as it is required for the cell membrane targeting of phosphofructokinase, an enzyme that catalyzes a rate-limiting reaction in glycolysis. [50]

In vitro studies have shown that Cav-3 plays a critical role in myoblast cell differentiation and survival and in myotube formation. [51] The relevance of Cav-3 in muscle physiology was further confirmed by the findings that mutations in the CAV3 gene result in distinct neuromuscular and cardiac disorders, such as limb girdle muscular dystrophy (LGMD) 1-C, idiopathic persistent elevation of serum creatine kinase (hyperCKemia), inherited rippling muscle disease (RMD), distal myopathy and familial hypertrophic cardiomyopathy (HCM). [52-54]

### CAVEOLAE AND CARDIAC ION CHANNELS

Modulation of ion channel activity plays a critical role in regulating cardiovascular function. Recently, it has become apparent that the regulation of channel function is not the only means of controlling excitability, the trafficking and localization of ion channels with signaling molecules also play a significant role. Most cells in the cardiovascular system express multiple channel types (e.g., voltage-gated  $Na^+$ ,  $K^+$  and  $Ca^{2+}$  channels) and even multiple isoforms of a particular channel, with each channel uniquely contributing to excitability. [55,56] Voltage gated

$Na^+$  channels are responsible for the initial depolarization of the cardiac sarcolemma, to permit the opening of voltage-gated L-type  $Ca^{2+}$  channels, resulting in  $Ca^{2+}$  influx and contraction. Membrane repolarization is controlled by  $K^+$  channels. Therefore, altering the number of channels and/or their function can have significant impact on both resting membrane potential and the cardiac action potential wave form. Defects in either of these processes can have life-threatening implications. [55,56]

In several cell types, including smooth muscle and endothelial cells, mediators of calcium signaling, such as  $Ca^{2+}$ -ATPase, inositol-triphosphate receptor (IP3R),  $Ca^{2+}$  pumps and L-type  $Ca^{2+}$  channels, large conductance  $Ca^{2+}$  activated  $K^+$  channel, calmodulin and transient receptor potential (TRP) channels, localize in cholesterol-rich membrane domains. Such localization suggest that membrane raft and/or caveolae have a role in calcium handling and  $Ca^{2+}$  entry that control excitation-contraction of heart muscle. [57,58] TRP channels, in particular TRPC1, -3 and -4 are enriched in caveolae and caveolin-1 regulates the plasma membrane localization and function of TRP channels. [59] Current evidence indicates that caveolae regulate calcium entry and depletion of cholesterol by methyl- $\beta$ -cyclodextrin reduces colocalization of caveolin-1 and TRPC1 and redistribution of TRPC1, thus preventing  $Ca^{2+}$  influx. [60] Moreover,  $Na^+$  pump,  $Na/K$ -ATPase, contains two caveolin binding motifs and resides in caveolae in a number of cells, including smooth muscle cells and cardiomyocytes, thereby helping to maintain  $Na^+$  gradient. [61] Voltage gated  $K^+$  channels are also localized in caveolae and play an important role to maintaining cellular excitability. In fibroblast, the Kv 1.5 subunit colocalizes with caveolin-1, Kv 2.5 localizes with membrane raft and depletion of cholesterol with M $\beta$ CD redistributes and alters the function of  $K^+$  channel. [62] These findings imply that alteration of caveolae and/or caveolin by any disease or drug

treatments can shift the localization of the channels, thereby altering cellular excitability and functional activity.

### **CAVEOLAE AND CARDIOVASCULAR DISEASE**

Normal heart physiology and vascular function is frequently disrupted and thereby gives rise to a multitude of pathological states. In recent years, many researchers have found that both caveolins and caveolae play a role in the development of various human diseases, including coronary heart disease, hypertension, stroke and nervous system disorders. [63]

There is a vast literature about the roles of caveolae and caveolin in the regulation of many cellular processes in cultured cells and many investigators considered them as an essential platform of signaling molecules. However, in the past few years, development of animal models and usage of genetically altered mice have been instrumental in deciphering their physiological functions in vivo. Transgenic over expression of caveolin-1 or caveolin-3 in mice or targeted disruption of each of the caveolin gene locus in mice (Cav-1, Cav-2 and Cav-3 genes) has provided significant insight into the roles of caveolin and caveolae. [64] The potential role of caveolin in cardiovascular physiology has become apparent by the discovery of caveolin-1 and caveolin-3 KO mice and double knockout mice, which have cardiomyopathic phenotype. Caveolin-1 KO mice show complete ablation of the presence of the caveolae, cellular organelle, in the endothelium and fat. Similarly, caveolin-3 KO mice lack caveolae in cells that normally express this protein such as skeletal muscle, heart and diaphragm. Heart tissue is made up of different types of cells. Differentiated cardiomyocytes surrounded by a network of cardiac fibroblasts and endothelial cells and less abundant vascular smooth muscle cells. There is also a controversy regarding expression of caveolin isoforms in the heart muscle. It is

well known that cardiac myocytes express caveolin-3 and other cell types in the heart express caveolin-1 and caveolin-2. But recent studies provided the evidence of the existence of caveolin-1 in cardiomyocytes. [65]

Caveolae and their coat proteins, caveolins (Cav), have diverse effects on endothelial function, nitric oxide synthesis regulation, signal transduction, cholesterol metabolism, and apoptosis. Animal studies in Cav knockout mice demonstrate the vital role of these structural proteins on endothelial and vascular function. Genetic studies have proposed that beside neoplasia, Caves may play a role in the development of atherosclerosis, cardiomyopathy, long QT syndrome, pulmonary fibrosis, and muscular dystrophy. Ongoing research is needed to clarify the diagnostic and prognostic role of these novel proteins and to determine how the effects of Caves can translate into clinical medicine. [66]

### **Caveolin and Atherosclerosis**

Atherosclerosis is a disease of the blood vessel characterized by the development of an arterial occlusion containing lipid and cellular deposits. Caveolae and caveolins are believed to play an important role in the regulation of cellular signaling and transport of molecules among others. Experimental evidence indicates that caveolae and caveolins have the possibility to influencing atherogenesis in many ways. Caveolin-1 is a cholesterol-binding protein that can transport cholesterol from the endoplasmic reticulum (ER) to the plasma membrane. The major receptors for high-density lipoprotein, SR-B1, and a scavenger receptor for modified forms of LDL, CD36, can also reside in and signal in caveolae-type microdomains. [67] In addition, oxidized LDL can extract caveolae cholesterol, unlocalize eNOS, and impair NO release. [68] Conversely, blockade of HMG CoA reductase with statin-based drugs reduces caveolin levels and promotes eNOS activation. [69] This concept has been

validated in apolipoprotein E-deficient (ApoE<sup>-/-</sup>) mice where statin treatment decreases caveolin-1 expression and promotes NOS function in vivo.<sup>[70]</sup> However, to date, there are no data showing changes in caveolin-1 levels in atherosclerotic lesions from humans.<sup>[64]</sup>

To verify, if caveolin-1 influenced lesion progression in mice, Lisanti and his coworkers crossbred caveolin-1<sup>-/-</sup> mice with ApoE<sup>-/-</sup> mice that develop atheromas. Interestingly, the loss of caveolin-1 in the ApoE<sup>-/-</sup> mice resulted in a proatherogenic lipid profile, similar to that seen in CD36<sup>-/-</sup> mice bred to an ApoE background.<sup>[71,72]</sup> Surprisingly, despite a pro-atherogenic lipid profile, the loss of caveolin-1 reduced lesion burden by 80%, suggesting caveolin-1 regulated LDL-mediated vascular dysfunction, inflammation, and lesion progression. The authors suggested this may be caused by a decrease in stability of the scavenger receptor for oxidized or modified LDL, CD36 in macrophages, and an increase in endothelium-derived NO production, which would reduce vascular inflammation. These remarkable findings unequivocally support the importance of caveolin-1/caveolae in the pathogenesis of atherosclerosis.<sup>[64]</sup>

But the role of Cav expression in atherosclerotic disease is poorly understood and remains controversial. Interestingly, there is emerging evidence between low Cav-1 levels and the vulnerable plaque, which could potentially identify Cav-1 as a novel plaque biomarker.<sup>[66]</sup>

Endothelial dysfunction is crucial in the initiation of atherosclerosis, which is associated with a lack of nitric oxide. The endothelial NO synthase (eNOS) is responsible for constitutive synthesis of NO and inhibited by caveolin-1 (Cav1). Loss of Cav1 increased vascular lesion by enhancing neointimal proliferation. The combined loss of Cav1 and eNOS, compared to Cav1<sup>-/-</sup>, lowered intima formation, suggesting an increasing effect of eNOS in the absence of Cav1 on vascular lesion.<sup>[73]</sup>

Global deletion of CAV1 in mice results in insulin resistance and increases in atherogenic plasma lipids and cholesterol, but protects from diet-induced obesity and atherosclerosis. In this study the cellular dynamics of intestinal Cav1 were visualized in zebrafish and the metabolic contributions of CAV1 were determined with mice lacking CAV1 in intestinal epithelial cells.<sup>[74]</sup>

### **Caveolin and Cardiac Hypertrophy**

The heart responds to multiple forms of stress with an adaptive hypertrophic increase in cardiac mass. Under prolonged stress, the heart undergoes an apparent irreversible change, resulting in dilation, diminished performance, and ultimate failure. Given that cardiac failure is the most common result of insufficiency of myocardium, it is not surprising that cardiomyocyte hypertrophy is the dominant cellular response to virtually all forms of hemodynamic overload.<sup>[75]</sup> However, long-term adaptive/compensatory hypertrophy is associated with progressive ventricular dilation. As a consequence of cardiac enlargement and wall thinning, stress on the wall also increases, despite constant intracavitary pressure. This mathematical increase in wall stress generates its own hemodynamic stress on the heart, further stimulating overloaded hypertrophy signaling pathway and thereby altering the balance from cell growth response to cell death. Once these processes have progressed to this stage (decompensation, loss of cardiac myocytes), irreversible functional deterioration develops, which leads to heart failure and, ultimately, death.<sup>[76,77]</sup>

Over-expression of caveolin-3 in neonatal cardiac myocytes decreases the ability of the adrenergic agonist phenylephrine or endothelin-1 to increase cell size.<sup>[74]</sup> A similar kind of effect is seen in cardiac myoblasts (H9C2) in which cav-3 reduces angiotensin II-promoted hypertrophy.<sup>[78]</sup> Other studies indicate that cardiac hypertrophy results in decreased expression of cav-3 and hypertrophy is enhanced in caveolin-1 KO and caveolin-

1/3 double KO mice. [79,80] Down regulation of growth signals are the most likely cause of expressed caveolin induced inhibition of cardiomyocyte growth. Cav-1 and -3 KO mice show hyperactivation of p42/44 MAPK [81] and upregulation of eNOS activity and nitrosative stress [82, 65] By contrast, increased caveolin expression down regulates activity of those entities. [83,84] Chronic myocardial hypoxia increases eNOS expression while decreasing the expression of cav-3, consistent with the idea that the expression and activity of eNOS is dependent on caveolin. [85] A recent finding indicate that caveolin-1 overexpression reduces hypertrophy by inhibiting autophagy pathway. [86] Alterations in caveolin expression almost certainly change the ability of the hypertrophied heart to respond to a variety of physiologic and pharmacologic agonists/ stimulus. [65]

#### **Caveolin and Myocardial Ischemia**

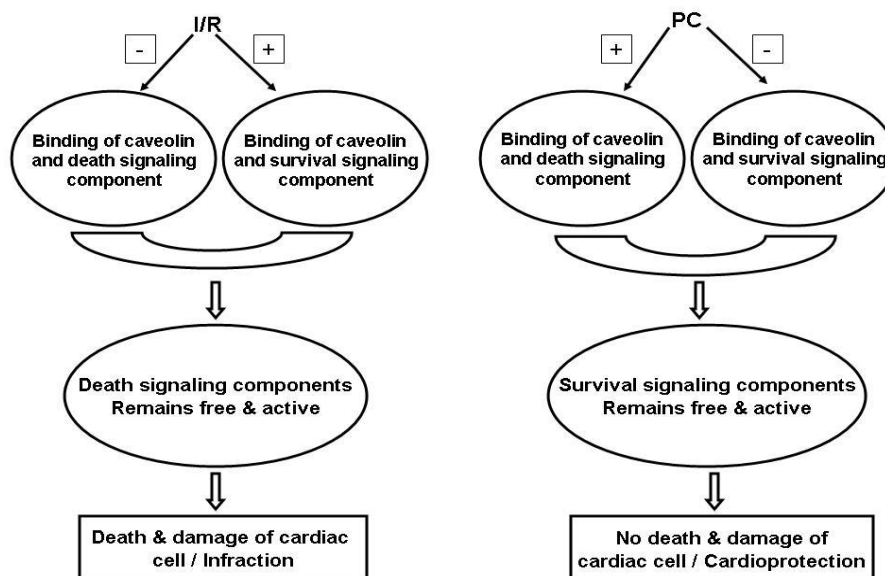
Myocardial infarction (i.e., heart attack) is the irreversible death (necrosis) of heart muscle secondary to prolonged lack of oxygen supply (ischemia). Ischemic heart disease is leading cause of death and disability worldwide. Precondition (PC) is the phenomenon whereby brief episodes of ischemia and reperfusion render the heart resistant to ischemic injury from a subsequent ischemic insult. Thus, ischemic PC is a protective and adaptive mechanism produced by short periods of ischemic stress rendering the heart more protected against another similar or greater stress. Early preconditioning depends on adenosine, opioids and to a lesser degree, on bradykinin and prostaglandins, released during ischemia. This molecule activate G-protein coupled receptor, initiates activation of  $K_{ATP}$  channel and generate oxygen free radicals, and stimulate a series of protein kinases, which include protein kinase C, tyrosine kinase and members of MAP kinase family. Late preconditioning is triggered by a similar sequence of events, but in addition essentially depends on newly synthesized proteins, which comprise iNOS, COX-2, manganese superoxide dismutase

and possibly heat shock proteins. The final mechanism of PC is still not very clear. However, evidence is rapidly accumulating about the involvement of caveolin or caveolae in cardioprotection against myocardial ischemia and ischemia/reperfusion injury. [87]

Ischemia/reperfusion injury activates p42/44 and p38MAPK, redistributes caveolin-3 and downregulates expression of caveolin-1. [87] Disruption of caveolae using M $\beta$ CD eliminates the ability of ischemia and pharmacological preconditioning to protect the cardiac myocyte from injury. [88] This is also supported by the decreased ability of Cav-1 KO mice to undergo pharmacological preconditioning. [87] Emerging evidences indicate that caveolin-1 (Cav-1), and caveolin-3 (Cav-3) both are essential for the protective effects of conditioning against myocardial I/R injury. [89] Recent investigations also showed that pro-survival signaling components translocate and/or interact with caveolin in ischemia/reperfusion heart and render the heart less abundance to pro-survival signal and induces myocardial injury. Similarly, in preconditioned heart death signaling components translocates and/or interact with caveolin in preconditioned heart and rendering the heart less exposed to death signaling components and more abundant to pro-survival signaling components [90] [Fig-2]. Although detail mechanism of action of caveolin is not very clear, but evidence indicates that proteasomes and histone modification play a very important role in the interaction between caveolin and signaling components. [91,92] We found that Cav-1 KO mouse abolished the acetylation of histone (H3 and H4) and increased the methylation of histone in the preconditioned heart. The increased histone methylation was significantly correlated with an increased level of histone methyltransferase G9a protein and increased the level of histone deacetylase (HDAC) activity. Recent investigation also shown that Cav-3 knockdown cells showed increased cell death and higher level of apoptotic proteins

(cleaved caspase-3 and cytochrome c) with suppressed mitochondrial function in response to simulated ischemia and I/R, whereas Cav-3 overexpressed cells were protected and had preserved mitochondrial function. [93] In the heart, autophagy may be a major regulator of protection from ischemic stress. It was found that Cav-3 knockdown cells have a decreased

expression of autophagy markers [beclin-1, light chain (LC3-II)] after simulated ischemia and ischemia-reperfusion (I/R) compared with WT, whereas overexpressed cells showed increased expression. [93] However, overall observation indicates that caveolin plays a pivotal role in cardioprotection against ischemic injury.



**Fig -2:** The role of caveolae in the ischemic preconditioning of the heart. In I/R heart, survival signaling components remain bound (+) with caveolin, whereas there was reduced association (-) of death signaling components with caveolin. These unbound death signaling components induces reperfusion injury in the heart. In PC heart, death signaling components remain bound (+) with caveolin, whereas there was reduced association (-) of survival signaling components with caveolin. These unbound anti-death/survival signaling components induced cardioprotection.

## CONCLUSION

Caveolae and caveolins are comparatively new players in a relatively saturated field of cardiovascular diseases and are undoubtedly regulating various aspects of cardiovascular system. Clearly loss of caveolin-1 has profound effect on the eNOS pathway, indicating the importance of this interaction, whereas the loss of caveolin-3 impacts NOS as well as MAPK activation and histone acetylation. Transgenic over expression of caveolin-1 or caveolin-3 in mice or targeted disruption of each of the caveolin gene locus in mice (Cav-1, Cav-2 and Cav-3 genes) has provided significant insight into the roles of caveolin and caveolae. The potential role of caveolin in cardiovascular physiology has become apparent by the discovery of caveolin-1 and caveolin-3 KO mice and

double knockout mice, which have cardiomyopathic phenotype. Although detail mechanisms of actions are not very clear, experimental evidences demonstrate the predominant role of caveolin in cardiac hypertrophy, atherosclerosis, ischemic injury and different myocardial functions. Recent investigations are disentangling the complex processes of caveolin regulated signaling systems in the myocardium and developing novel approaches, aimed at counteracting cardiomyocyte apoptosis in heart failure and/or cardiovascular diseases.

## REFERENCES

1. Michel V, Bakovic M (2007) Lipid rafts in health and disease. *Biol Cell.* 99: 129–40.
2. Lisanti MP, Scherer PE, Tang Z, Sargiacomo M (1994) Caveolae,



- caveolin and caveolin-rich membrane domains: A signalling hypothesis. *Trends Cell Biol.* 4:231–235.
3. Ostrom RS, Gregorian C, Drenan RM, Xiang Y, Regan JW, Insel PA (2001) Receptor number and caveolar co-localization determine receptor coupling efficiency to adenylyl cyclase. *J Biol Chem* 276: 42063–42069.
  4. Anderson RG (1993) Potocytosis of small molecules and ions by caveolae. *Trends Cell Biol.* 3 :69–72
  5. Fujimoto T (1993) Calcium pump of the plasma membrane is localized in caveolae. *J Cell Biol.* 120: 1147–1157.
  6. Horikawa YT, Tsutsumi YM, Patel HH, Roth DM (2014) Signaling epicenters: the role of caveolae and caveolins in volatile anesthetic induced cardiac protection. *Curr Pharm Des* 20(36): 5681-5689
  7. Suzuki Y, Yamamura H, Ohya S, Imaizumi Y(2013) Direct molecular interaction of caveolin-3 with KCa1.1 channel in living HEK293 cell expression system. *Biochemical and biophysical research communications.* 430:1169–1174
  8. Chiari PC, Pagel PS, Tanaka K, Krolkowski JG, Ludwig LM, Trillo RA, Jr, Puri N, Kersten JR, Warltier DC (2004) Intravenous emulsified halogenated anesthetics produce acute and delayed preconditioning against myocardial infarction in rabbits. *Anesthesiology.* 101:1160–1166
  9. Wyse BD, Prior IA, Qian H (2003) Caveolin interacts with the angiotensin II type 1 receptor during exocytic transport but not at the plasma membrane. *J Biol Chem.* 278: 23738–23746.
  10. Cohen AW, Hnasko R, Schubert W, Lisanti MP (2004) Role of caveolae and caveolins in health and disease. *Physiol Rev.* 84: 1341–1379.
  11. Insel PA, Patel HH (2007) Do studies in caveolin-knockouts teach us about physiology and pharmacology or instead the ways mice compensate for 'lost proteins'? *Br Pharmacol.* 150: 251–54.
  12. Lee H, Woodman SE, Engelman JA (2001) Palmitoylation of caveolin-1 at a single site (Cys-156) controls its coupling to the c-Src tyrosine kinase: targeting of dually acylated molecules (GPI-linked, transmembrane, or cytoplasmic) to caveolae effectively uncouples c-Src and caveolin-1 (TYR-14) *J Biol Chem.* 276: 35150–35158
  13. Parat MO, Fox PL (2001) Palmitoylation of caveolin-1 in endothelial cells is post-translational but irreversible. *J Biol Chem.* 276: 15776–82.
  14. Garcia-Cardena G, Fan R, Stern DF, Liu J, Sessa WC (1996) Endothelial nitric oxide synthase is regulated by tyrosine phosphorylation and interacts with caveolin-1. *J Biol Chem.* 271: 27237–27240.
  15. Venema VJ, Ju H, Zou R, Venema RC (1997) Interaction of neuronal nitric-oxide synthase with caveolin-3 in skeletal muscle. Identification of a novel caveolin scaffolding/inhibitory domain. *J Biol Chem.* 272: 28187–28190.
  16. Razani B, Engelman JA, Wang XB (2001) Caveolin-1 null mice are viable but show evidence of hyperproliferative and vascular abnormalities. *J Biol Chem.* 276: 38121–38138.
  17. Toya Y, Schwencke C, Couet J, Lisanti MP, Ishikawa Y (1998) Inhibition of adenylyl cyclase by caveolin peptides. *Endocrinology.* 139: 2025–2031
  18. Razani B, Lisanti MP (2001) Two distinct caveolin-1 domains mediate the functional interaction of caveolin-1 with protein kinase A. *Am J Physiol Cell Physiol.* 281: C1241–50
  19. Peart JN, Headrick JP (2007) Adenosinergic cardioprotection: multiple receptors, multiple pathways. *Pharmacol Ther.* 114: 208–221.
  20. Zhuang L, Lin J, Lu ML, Solomon KR, Freeman MR (2002) Cholesterol-rich lipid rafts mediate akt-regulated survival in prostate cancer cells. *Cancer Res.* 62: 2227–2231.
  21. Li L, Ren CH, Tahir SA, Ren C, Thompson TC (2003) Caveolin-1 maintains activated Akt in prostate cancer cells through scaffolding domain binding site interactions with and inhibition of serine/threonine protein phosphatases PP1 and PP2A. *Mol Cell Biol.* 23: 9389–9404.

22. Smythe GM, Rando TA (2006) Altered caveolin-3 expression disrupts PI(3) kinase signaling leading to death of cultured muscle cells. *Exp Cell Res.* 312: 2816–25.
23. Zhang B, Peng F, Wu D, Ingram AJ, Gao B, Krepinsky JC (2007) Caveolin-1 phosphorylation is required for stretch-induced EGFR and Akt activation in mesangial cells. *Cell Signal.* 19: 1690–1700.
24. Couet J, Sargiacomo M, Lisanti MP (1997) Interaction of a receptor tyrosine kinase, EGF-R, with caveolins. Caveolin binding negatively regulates tyrosine and serine/threonine kinase activities. *J Biol Chem.* 272: 30429–30438.
25. Pike LJ (2005) Growth factor receptors, lipid rafts and caveolae: an evolving story. *Biochim Biophys Acta.* 1746: 260–273.
26. Galbiati F, Volonte D, Engelman JA. (1998) Targeted downregulation of caveolin-1 is sufficient to drive cell transformation and hyperactivate the p42/44 MAP kinase cascade. *EMBO J.* 17: 6633–6648.
27. Engelman JA, Chu C, Lin A (1998) Caveolin-mediated regulation of signaling along the p42/44 MAP kinase cascade *in vivo*. A role for the caveolin-scaffolding domain. *FEBS Lett.* 428: 205–211.
28. Cohen AW, Park DS, Woodman SE (2003) Caveolin-1 null mice develop cardiac hypertrophy with hyperactivation of p42/44 MAP kinase in cardiac fibroblasts. *Am J Physiol Cell Physiol.* 284: C457–C474.
29. Ballard-Croft C, Locklar AC, Kristo G, Lasley RD (2006) Regional myocardial ischemia-induced activation of MAPKs is associated with subcellular redistribution of caveolin and cholesterol. *Am J Physiol Heart Circ Physiol.* 291: H658–67
30. Song KS, Sargiacomo M, Galbiati F, Parenti M, Lisanti MP (1997) Targeting of a G $\alpha$  subunit (G $\alpha$ i1) and c-Src tyrosine kinase to caveolae membranes: clarifying the role of N-myristoylation. *Cell Mol Biol.* 43: 293–303.
31. Li S, Couet J, Lisanti MP (1996) Src tyrosine kinases, G $\alpha$  subunits, and H-Ras share a common membrane-anchored scaffolding protein, caveolin. Caveolin binding negatively regulates the auto-activation of Src tyrosine kinases. *J Biol Chem.* 271: 29182–29190.
32. Lee H, Volonte D, Galbiati F, Iyengar P, Lublin DM (2000) Constitutive and growth factor-regulated phosphorylation of caveolin-1 occurs at the same site (Tyr-14) *in vivo*: identification of a c-Src/Cav-1/Grb7 signaling cassette. *Mol Endocrinol.* 14: 1750–75.
33. Woodman SE, Park DS, Cohen AW, Cheung MW, Chandra M, Shirani J (2002) Caveolin-3 knockout mice develop a progressive cardiomyopathy and show hyperactivation of the p42/44 MAPK cascade. *J Biol Chem.* 277: 38988–38997.
34. Park DS, Woodman SE, Schubert W, Cohen AW, Frank PG, Chandra M (2002) Caveolin 1/3 double knockout mice are viable but lack both muscle and non-muscle caveolae and develop a severe cardiomyopathic phenotype. *Am J Pathol.* 160: 2207–2217.
35. Murata T, Lin MI, Huang Y, Yu J, Bauer PM, Giordano FJ, Sessa WC (2007) Reexpression of caveolin-1 in endothelium rescues the vascular, cardiac, and pulmonary defects in global caveolin-1 knockout mice. *J Exp Med.* 204(10): 2373–2382.
36. Wunderlich C, Schober K, Lange SA (2006) Disruption of caveolin-1 leads to enhanced nitrosative stress and severe systolic and diastolic heart failure. *Biochem Biophys Res Commun.* 340(2):702–708.
37. Wang W, Schulze CJ, Suarez-Pinzon WL, Dyck JR, Sawicki G, Schulz R (2002) Intracellular action of matrix metalloproteinase-2 accounts for acute myocardial ischemia and reperfusion injury. *Circulation.* 106: 1543–1549.
38. Sawicki H, Leon H, Sawicka J (2005) Degradation of myosin light chain in isolated rat hearts subjected to ischemia-reperfusion injury: a new intracellular target for matrix metalloproteinase-2. *Circulation* 112: 544–552
39. Nagase H, Visse R, Murphy (2006) Structure and function of matrix

- metalloproteinases and TIMPs. *Cardiovasc Res.* 69: 562–573.
40. Puyraimond A, Fridman R, Lemesle M, Arbeille B, Menashi (2001) MMP-2 colocalizes with caveolae on the surface of endothelial cells. *Exp Cell Res.* 262: 28–36.
  41. Atkinson SJ, English JL, Holway N, Murphy G (2004) Cellular cholesterol regulates MT1 MMP dependent activation of MMP 2 *via* MEK-1 in HT1080 fibrosarcoma cells. *FEBS Lett.* 566: 65–70.
  42. Fiucci G, Ravid D, Reich R, Liscovitch M.(2002) Caveolin-1 inhibits anchorage-independent growth, anoikis and invasiveness in MCF-7 human breast cancer cells. *Oncogene*21:2365–2375.
  43. Woodman SE, Park DS, Cohen AW (2002) Caveolin-3 knock-out mice develop a progressive cardiomyopathy and show hyperactivation of the p42/44 MAPK cascade. *J Biol Chem.* 277(41):38988–38997.
  44. Capozza F, Cohen AW, Cheung MW (2005) Muscle-specific interaction of caveolin isoforms: differential complex formation between caveolins in fibroblastic vs. muscle cells. *Am J Physiol Cell Physiol.* 288: C677–C691
  45. Song KS, Scherer PE, Tang Z (1996) Expression of caveolin-3 in skeletal, cardiac, and smooth muscle cells. Caveolin-3 is a component of the sarcolemma and co-fractionates with dystrophin and dystrophin-associated glycoproteins. *J Biol Chem.* 271:15160–15165
  46. Crosbie RH, Yamada H, Venzke DP (1998) Caveolin-3 is not an integral component of the dystrophin glycoprotein complex. *FEBS Lett.*427:279–82.
  47. Sotgia F, Lee JK, Das K (2000) Caveolin-3 directly interacts with the C-terminal tail of beta—dystroglycan. Identification of a central WW-like domain within caveolin family members. *J Biol Chem.* 275: 38048–38058.
  48. Garcia-Cardena G, Fan R, Stern DF (1996) Endothelial nitric oxide synthase is regulated by tyrosine phosphorylation and interacts with caveolin-1. *J Biol Chem.* 271: 27237–27240.
  49. Smythe GM, Eby JC, Disatnik MH (2003) A caveolin-3 mutant that causes limb girdle muscular dystrophy type 1C disrupts Src localization and activity and induces apoptosis in skeletal myotubes. *J Cell Sci.* 116: 4739–4749.
  50. Sotgia F, Bonuccelli G, Minetti C (2003) Phosphofructokinase muscle-specific isoform requires caveolin-3 expression for plasma membrane recruitment and caveolar targeting: implications for the pathogenesis of caveolin-related muscle diseases. *Am J Pathol.* 163: 2619–2634.
  51. Galbiati F, Volonte D, Engelman JA (1999) Targeted down-regulation of caveolin-3 is sufficient to inhibit myotube formation in differentiating C2C12 myoblasts. Transient activation of p38 mitogen-activated protein kinase is required for induction of caveolin-3 expression and subsequent myotube formation. *J Biol Chem.* 274: 30315–30321.
  52. Minetti C, Sotgia F, Bruno C (1998) Mutations in the caveolin-3 gene cause autosomal dominant limb-girdle muscular dystrophy. *Nat Genet.* 18: 365–368.
  53. Carbone I, Bruno C, Sotgia F (2000) Mutation in the CAV3 gene causes partial caveolin-3 deficiency and hyperCKemia. *Neurology.* 54: 1373–1376.
  54. Galbiati F, Razani B, Lisanti MP (2001) Caveolae and caveolin-3 in muscular dystrophy. *Trends Mol Med.* 7: 435–441.
  55. O'Connell KM, Martens JR, Tamkun MM.(2004) Localization of ion channels to lipid Raft domains within the cardiovascular system. *Trends Cardiovasc Med.* 14(2): 37–42.
  56. Maguy A, Hebert TE, Nattel S (2006) Involvement of lipid rafts and caveolae in cardiac ion channel function. *Cardiovasc Res.* 69(4): 798–807.
  57. Fujimoto T, Miyawaki A, Mikoshiba K (1995) Inositol 1,4,5-trisphosphate receptor-like protein in plasmalemma caveolae is linked to actin filaments. *J Cell Sci.* 108((Pt. 1)):7–15.
  58. Wang XL, Ye D, Peterson TE (2005) Caveolae targeting and regulation of

- large conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels in vascular endothelial cells. *J Biol Chem.* 280: 11656–11664.
59. Kwiatek AM, Minshall RD, Cool DR, Skidgel RA, Malik AB, Tiruppathi C (2006) Caveolin-1 regulates store-operated  $\text{Ca}^{2+}$  influx by binding of its scaffolding domain to transient receptor potential channel-1 in endothelial cells. *Mol Pharmacol.* 70: 1174–1183.
60. Bergdahl A, Gomez MF, Dreja K (2003) Cholesterol depletion impairs vascular reactivity to endothelin-1 by reducing store-operated  $\text{Ca}^{2+}$  entry dependent on TRPC1. *Circ Res.* 93: 839–847.
61. Fagan KA, Smith KE, Cooper DM (2000) Regulation of the  $\text{Ca}^{2+}$ -inhibitable adenylyl cyclase type VI by capacitative  $\text{Ca}^{2+}$  entry requires localization in cholesterol-rich domains. *J Biol Chem.* 275: 26530–26537.
62. Martens JR, Sakamoto N, Sullivan SA, Grobaski TD, Tamkun MM (2001) Isoform-specific localization of voltage-gated  $\text{K}^+$  channels to distinct lipid raft populations. Targeting of Kv1.5 to caveolae. *J Biol Chem.* 276: 8409–8414.
63. Yin H, Liu T, Zhang Y, Yang B (2016) Caveolin proteins: a molecular insight into disease. *Front Med.*10(4):397-404.
64. Gratton JP, Bernatchez P, Sessa WC (2008) Caveolae and caveolins in the cardiovascular system. *Circ Res.* 2004;94(11):1408–17.
65. Patel HH, Murray F, Insel PA (2008) Caveolae as organizers of pharmacologically relevant signal transduction molecules. *Annu Rev Pharmacol Toxicol.* 48: 359–391.
66. Sanon VP, Sawaki D, Mjaatvedt CH, (2015) Myocardial tissue caveolae. *Compr Physiol.* 5(2):871-86.
67. Graf GA, Matveev SV, Smart EJ (1999) Class B scavenger receptors, caveolae and cholesterol homeostasis. *Trends Cardiovasc Med.* 9: 221–225.
68. Blair A, Shaul PW, Yuhanna IS, Conrad PA, Smart EJ (1999) Oxidized low density lipoprotein displaces endothelial nitric-oxide synthase (eNOS) from plasmalemmal caveolae and impairs eNOS activation. *J Biol Chem.* 274: 32512–32519.
69. Feron O, Dessy C, Desager JP, Balligand JL (2001) Hydroxymethylglutaryl-coenzyme a reductase inhibition promotes endothelial nitric oxide synthase activation through a decrease in caveolin abundance. *Circulation.* 103: 113–118.
70. Pelat M, Dessy C, Massion P, Desager JP, Feron O, Balligand JL (2003) Rosuvastatin decreases caveolin-1 and improves nitric oxide-dependent heart rate and blood pressure variability in apolipoprotein E<sup>-/-</sup> mice *in vivo*. *Circulation.* 107: 2480–2486.
71. Febbraio M, Podrez EA, Smith JD (2000) Targeted disruption of the class B scavenger receptor CD36 protects against atherosclerotic lesion development in mice. *J Clin Invest.* 105: 1049–1056.
72. Podrez EA, Febbraio M, Sheibani N (2000) Macrophage scavenger receptor CD36 is the major receptor for LDL modified by monocyte-generated reactive nitrogen species. *J Clin Invest.* 105: 1095–1098.
73. Mierke J, Christoph M, Pfluecke C, Jellinghaus S, Wunderlich C, Strasser RH, Ibrahim K, Poitz DM.(2017) Atheroprotective role of Caveolin-1 and eNOS in an innovative transplantation model is mainly mediated by local effects. *Biochim Biophys Acta.*1863(2):529-536
74. Otis JP, Shen MC, Quinlivan V, Anderson JL, Farber SA. (2017) Intestinal epithelial cell caveolin 1 regulates fatty acid and lipoprotein cholesterol plasma levels. *Dis Model Mech.*10(3):283-295
75. Diwan A, Dorn GW (2007) Decompensation of cardiac hypertrophy: cellular mechanism and novel therapeutic target. *Physiology.* 22: 56–64
76. Hill JA, Karimi M, Kutschke W (2000) Cardiac hypertrophy is not a required compensatory response to short term pressure overload. *Circulation.* 101: 2863–2869.
77. Sano M, Schneider MD (2002) Still stressed out but doing fine: normalization of wall stress is superfluous to maintain cardiac function

- in chronic pressure overload. *Circulation*. 105: 8–10.
78. Koga A, Oka N, Kikuchi T, Miyazaki H, Kato S, Imaizumi T (2003) Adenovirus-mediated overexpression of caveolin-3 inhibits rat cardiomyocyte hypertrophy. *Hypertension*. 42: 213–219.
79. Fujita T, Otsu K, Oshikawa J (2006) Caveolin-3 inhibits growth signal in cardiac myoblasts in a Ca<sup>2+</sup>-dependent manner. *J Cell Mol Med*. 10: 216–224.
80. De Souza AP, Cohen AW, Park DS (2005) MR imaging of caveolin gene-specific alterations in right ventricular wall thickness. *Magn Reson Imaging*. 23: 61–68.
81. Wunderlich C, Schober K, Lange SA (2006) Disruption of caveolin-1 leads to enhanced nitrosative stress and severe systolic and diastolic heart failure. *Biochem Biophys Res Commun*. 340: 702–708.
82. Woodman SE, Park DS, Cohen AW (2002) Caveolin-3 knock-out mice develop a progressive cardiomyopathy and show hyperactivation of the p42/44 MAPK cascade. *J Biol Chem*. 277: 38988–38997.
83. Feron O, Balligand JL (2006) Caveolins and the regulation of endothelial nitric oxide synthase in the heart. *Cardiovasc Res*. 69: 788–797.
84. Garcia-Cardena G, Martasek P, Masters BS (1997) Dissecting the interaction between nitric oxide synthase (NOS) and caveolin. Functional significance of the nos caveolin binding domain *in vivo*. *J Biol Chem*. 272: 25437–25440.
85. Patel HH, Tsutsumi YM, Head BP (2007) Mechanisms of cardiac protection from ischemia/reperfusion injury: a role for caveolae and caveolin-1. *FASEB J*. 21(7):1565–1574.
86. Wu D, Xie F, Xiao L, Feng F, Huang S, He L, Liu M, Zhou Q, Li L, Chen L. (2017). Caveolin-1-Autophagy Pathway Mediated Cardiomyocyte Hypertrophy Induced by Apelin-13. *DNA Cell Biol*. 2017 May 24. doi: 10.1089/dna.2016.3574.
87. Ballard-Croft C, Locklar AC, Kristo G, Lasley RD (2006) Regional myocardial ischemia-induced activation of MAPKs is associated with subcellular redistribution of caveolin and cholesterol. *Am J Physiol Heart Circ Physiol*. 291: H658–H667.
88. Das M, Gherghiceanu M, Lekli I, Mukherjee S, Popescu LM, Das DK (2008) Essential role of lipid raft in ischemic preconditioning. *Cell Physiol Biochem*. 21(4): 325–334.
89. Yang Y, Ma Z, Hu W, Wang D, Jiang S, Fan C, Di S, Liu D, Sun Y, Yi W (2016). Caveolin-1/-3: therapeutic targets for myocardial ischemia/reperfusion injury. *Basic Res Cardiol*. 111(4):45
90. Das M, Cui J, Das DK (2007) Generation of survival signal by differential interaction of p38MAPKalpha and p38MAPKbeta with caveolin-1 and caveolin-3 in the adapted heart. *J Mol Cell Cardiol*. 42(1):206–213.
91. Das M, Das S, Wang P, Powell SR, Das DK (2008) Caveolin and proteasome in tocotrienol mediated myocardial protection. *Cell Physiol Biochem*. 22(1-4): 287-294.
92. Das M, Das S, Lekli I, Das DK (2012) Caveolin induces cardioprotection through epigenetic regulation. *J Cell Mol Med*. 16(4): 888-895.
93. Kassan A, Pham U, Nguyen Q, Reichelt ME, Cho E, Patel PM, Roth DM, Head BP, Patel HH. (2016) Caveolin-3 plays a critical role in autophagy after ischemia-reperfusion. *Am J Physiol Cell Physiol*. 311(6):C854-C865.

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