

Original Research Article

## In Vitro Activity of Tigecycline against Multidrug Resistant Gram Negative Bacilli as Evaluated by Disc Diffusion Method and E-Test

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Received: 15/11/2013

Revised: 18/12/2013

Accepted: 19/12/2013

### ABSTRACT

**Introduction:** Tigecycline is an antibiotic belonging to the glycyclines class with *in vitro* activity against most gram negative bacteria, even multidrug resistant pathogens. It is considered to be a newer treatment option for emerging multidrug resistant pathogens.

**Objectives:** To evaluate the *in vitro* activity of tigecycline against Multidrug resistant gram negative bacteria isolated from various clinical specimens to compare with other antimicrobials.

**Materials & methods:** A total of 150 multidrug resistant isolates of Enterobacteriaceae (113) and Acinetobacter spp (37) were tested for tigecycline susceptibility by the E-test and disc diffusion method.

**Results:** Tigecycline showed good microbiological activity against all the isolates of multidrug resistant gram negative bacteria with 100% susceptibility in E. coli & Enterobacter species, 94% in Klebsiella species and 81.08% in Acinetobacter spp. isolates.

**Conclusion:** Tigecycline was found to be highly effective against multidrug resistant gram negative bacteria. Therefore it is an alternative option for treatment of complicated skin & soft tissue and intra-abdominal infections caused by such multidrug resistant pathogens.

**Key words:** Tigecycline, multidrug resistance (MDR), *in vitro* susceptibility, ESBL, MBL

### INTRODUCTION

The rates of antimicrobial drug resistance and particularly of multiple drug resistance are increasing among gram negative organisms, thus posing a difficult challenge to treat such infections. <sup>(1)</sup> Multi drug resistance in clinically important organisms particularly pathogens of family that produce  $\beta$ -lactamases with a broad

profile of substrate activity such as extended-spectrum  $\beta$ -lactamases (ESBLs), AmpC  $\beta$ -lactamases, as well as carbapenemases, including metallo  $\beta$ -lactamases (MBLs) and non-fermentative gram negative bacilli (including Acinetobacter spp & Pseudomonas spp) have led to the limited therapeutic options,

resulting in increased morbidity and mortality. (2,3)

Patient will respond to antibiotic if the pathogen is susceptible to the chosen antibiotic; however today situation is so worrisome that no agents available that are fully active against all the common pathogens. The key to antimicrobial development has been to design agents that elude the main bacterial resistance mechanisms. One such agent is Tigecycline, which is chemically the 9-t-butylglycylamido derivative of minocycline, is a member of a novel class of antibiotics, the glycylcyclines. Like the tetracyclines, tigecycline binds to the 30s subunit of bacterial ribosomes and inhibits protein synthesis by preventing the incorporation of aminoacid residues into elongating peptide chains. (4,5)

Regarding Enterobacteriaceae, tigecycline has shown to evade common mechanisms of acquired tetracycline resistance, such as those conferred by efflux pumps encoded by the (A-D) resistance determinants and ribosomal protective mechanisms. (6) However it has been reported that tigecycline showed only bacteriostatic activity against bacterial isolates of Acinetobacter species. (7,8) Nevertheless, tigecycline clearly displays inhibitory activity against Acinetobacter spp. (9-12) and has been utilized for therapy against MDR strains despite the lack of US FDA approved clinical indication & interpretative criteria for in vitro susceptibility testing. (13)

The drug is not significantly active against *Pseudomonas aeruginosa* & *Proteaeae* as it carry inherently encoded resistance-nodulation-division (RND) efflux pumps that confer decreased sensitivity. (14,15)

The drug was approved for use by the US Food and Drug Administration (FDA) in June 2005 and by European medicine agency in April 2006 for empiric monotherapy of nosocomial and acquired

intra-abdominal infection (IAI) and skin and soft tissue infections. Recently in 2009, the US Food and Drug Administration (FDA) approved tigecycline for community-acquired pneumonia. (16)

Therefore considering increasing rate of MDR gram negative pathogens, we evaluated the in-vitro activity of tigecycline against multiple-drug resistant *E. coli*, *Klebsiella*, *Enterobacter* and *Acinetobacter* spp. and compared its activity against other commonly used antibiotics.

## MATERIALS & METHODS

A total of 150 Gram negative isolates included in this study were isolated from patients attended to a tertiary care teaching hospital in North Maharashtra. Among these 150 isolates, 29 were *E. coli* spp, 50 were *Klebsiella* spp, 34 were *Enterobacter* spp and 37 were *Acinetobacter* spp. All isolates were selected from an existing stock of organisms isolated retrospectively over 2 year period starting from January 2011. Only one isolate per patient was included for testing.

All the test strains were isolated and identified by conventional biochemical tests. (17)

Antimicrobial susceptibility testing was done by Kirby-baur disc diffusion method as per the Clinical and Laboratory Standard Institute (CLSI) guidelines (18) using the Mueller Hinton agar and antimicrobial discs. The following antimicrobial agents ( $\mu\text{g}$ ) were used.

Amikacin (30  $\mu\text{g}$ ), amoxicillin/clavulanic acid (20/10 $\mu\text{g}$ ), aztreonam (30  $\mu\text{g}$ ), ceftazidime (30  $\mu\text{g}$ ), ciprofloxacin (5  $\mu\text{g}$ ), cefepime (30  $\mu\text{g}$ ), gentamicin(10  $\mu\text{g}$ ), imipenem (10 $\mu\text{g}$ ), meropenem (10 $\mu\text{g}$ ), piperacillin/tazobactam(100/10 $\mu\text{g}$ ), trimethoprim-sulphamethoxazole (25 $\mu\text{g}$ ) and tigecycline (15  $\mu\text{g}$ ).

The presence of ESBL in isolates of Enterobacteriaceae was screened by double

disc approximation test using ceftazidime and ceftazidime-clavulanic acid discs according to CLSI guidelines (18) and confirmed by E test. E.coli ATCC 25922 and Klebsiella pneumoniae ATCC 700603 were used as negative and positive controls respectively for testing ESBL production.

Isolates of Acinetobacter species were screened for MBL production using imipenem and imipenem-EDTA combined disc diffusion method (19) and confirmed by E-test.

In this study, isolates were defined as multi-drug resistant when they demonstrated diminished susceptibility to  $\geq 2$  of drug classes tested in susceptibility testing panel. (20)

Tigecycline susceptibility screening was initially done by disc diffusion method using tigecycline disc (15µg). Tigecycline MIC was determined using the E-test according to manufacturer's instructions & CLSI guidelines. (18) All the antibiotic discs, media, E strips and ATCC strains were supplied by Himedia laboratories, Mumbai. Interpretation of the Antimicrobial susceptibility testing was done as per CLSI

criteria. Since there were no CLSI recommended interpretative criteria for tigecycline, the US FDA breakpoints: Enterobacteriaceae (susceptible when MIC  $\leq 2$  µg/ml and resistant  $\geq 8$  µg/ml) were used.

## RESULTS

A total of 150 MDR gram negative isolates were evaluated in this study. These included 113 of Enterobacteriaceae and 37 of Acinetobacter spp. The source of these isolates included pus (51), blood (9), Respiratory samples (29), urine (27), sterile body fluids (8), wound swabs (21) and other specimens (ear swabs, skin swabs - 5).

Since tigecycline has no or limited activity against Pseudomonas and Proteus species, (21) these were not included in this study.

Ninety six percent of multidrug resistant Enterobacteriaceae strains were positive for ESBL production and 67.56% of multidrug resistant Acinetobacter spp were positive for MBL production. The complete antibiotic susceptibility profile of the tested organism is given in Table-1.

Table 1: Antibiotic susceptibility for MDR GNB.

Name of Antibiotic	Enterobacteriaceae (113) {E. coli (29), Klebsiella (50), Enterobacter (34)}			Acinetobacter (37)		
	Sensitive	Intermediate	Resistant	Sensitive	Intermediate	Resistant
Amikacin	71	19	23	15	00	22
Amoxicillin-clav	5	7	101	12	07	18
Aztreonam	5	6	102	3	7	27
Ceftazidime	00	07	106	4	0	33
Cefepime	42	07	64	5	00	32
Ciprofloxacin	11	08	94	1	2	34
Gentamicin	34	02	97	9	0	28
Imipenem	108	01	04	22	01	14
Meropenem	108	03	02	25	00	12
Piperacillin-tazobactam	57	25	31	17	05	15
Tigecyclin	113	00	00	31	02	04
Trimethoprim-sulfamethoxazole	15	02	96	05	00	32

Table 2 shows the activity of tigecycline against MDR Gram negative isolates tested by E test method.

Organism (150)	Susceptible (%)	Intermediate (%)	Resistant (%)
E.coli (29)	29 (100)	-	-
Klebsiella spp (50)	47 (94)	2 (4)	1(2)
Enterobacter spp (34)	34 (100)	-	-
Acinetobacter spp (37)	30 (81.08)	2 (5.4)	5 (13.5)

Table 3- Distribution of tigecycline MICs against MDR gram negative isolates.

Organism (No.)	MIC50	MIC90	MIC range
E. coli (29)	0.25	0.50	0.047-6
Klebsiella (50)	0.50	2	0.25-16
Enterobacter (34)	0.25	0.50	0.047-8
Acinetobacter (37)	1	4	0.25-16

Thus when evaluated by disk-diffusion method, tigecycline was 100% active against isolates of *E. coli*, *Klebsiella* species and *Enterobacter* species. It was also active against 86.5% of *Acinetobacter* species.

Activity of tigecycline against MDR gram negative isolated tested by E-test is shown in Table-2 and MIC values are given in Table-3.

There are currently no interpretative MIC breakpoints available from the CLSI for tigecycline. Based on breakpoints recommended by US Food and Drug Administration for Enterobacteriaceae (susceptible  $\leq 2\text{mg/l}$ , resistant  $\geq 8\text{mg/l}$ ), tigecycline was found to be 100% active against *E. coli* and *Enterobacter* spp. It was also active against 94% of *Klebsiella* spp.

At present, there are no interpretative breakpoints available for *Acinetobacter* species. If the same interpretative criteria for Enterobacteriaceae are arbitrarily applied, 81.08 % of the tested *Acinetobacter* species were susceptible.

The MIC 50 & MIC 90 for *E. coli* and *Enterobacter* species in this study was 0.25 and 0.5  $\mu\text{g/ml}$ . Diameter of the zone of inhibition for tigecycline in these isolates ranged between 23-30 mm and MIC range was 0.047- 6  $\mu\text{g/ml}$  for *E. coli* and 0.047-8  $\mu\text{g/ml}$  for *Enterobacter* species.

MIC 50 & MIC 90 for MDR *Klebsiella* species was 0.50 $\mu\text{g/ml}$  & 2 $\mu\text{g/ml}$  respectively. Diameter of the zone of inhibition for tigecycline in these isolates ranged between 17-30 mm, and MIC range was 0.25-16  $\mu\text{g/ml}$  for *Klebsiella* spp.

MIC 50 & MIC 90 for MDR *Acinetobacter* species was 0.25 $\mu\text{g/ml}$  & 16 $\mu\text{g/ml}$  respectively. Four out of 37 isolates had MIC values of  $\geq 8 \mu\text{g/ml}$  i.e. in resistant range and all four showed  $\leq 19$  mm zone diameter by disc diffusion method i.e. resistant . Five isolates had MIC values in

the intermediate range (3-8  $\mu\text{g/ml}$ ). Out of five isolates with MIC values of intermediate range, only two were resistant by disc diffusion method ( $\leq 19$  mm).

## DISCUSSION

Tigecycline has broad spectrum in-vitro activity against gram positive, gram negative and anaerobes. In addition, tigecycline demonstrates in-vitro activity against MRSA, VRE & ESBL producing pathogens. Also tigecycline has promising microbiological, pharmacodynamics & pharmacokinetic profile, therefore it is considered as a good alternative to treat infections due to multidrug resistant organisms. <sup>(22)</sup>

In present study, all the multidrug resistant *E. coli* and *Enterobacter* species isolates were found to be sensitive to tigecycline. MIC 50 & MIC 90 values of tigecycline were 0.25 $\mu\text{g/ml}$  and 0.50  $\mu\text{g/ml}$ , which correlates with findings of previous Indian studies<sup>(23-25)</sup> and foreign studies.<sup>(26-28)</sup>

The susceptibility to tigecycline of *Klebsiella* species in present study was 94%, which correlates with findings in studies done by Souli M et al <sup>(26)</sup> who reported 92.6% and by Ralf et al <sup>(29)</sup> who reported 92.5% cumulative susceptibility rate of tigecycline in ESBL *Klebsiella* species. In the systematic review by Theodoros Kelesidis, after evaluating 23 studies it was found that cumulative susceptibility rate to tigecycline of multidrug resistant *Klebsiella* species was 91.2% for 2627 isolates. <sup>(21)</sup>

In this study, MIC50 & MIC90 values of tigecycline against *Acinetobacter* species determined by E-test were 1  $\mu\text{g/ml}$  & 4  $\mu\text{g/ml}$  respectively, and among 37 isolates tested, susceptibility to tigecycline was 81.08%.

In another study from India, 70.6% of MDR *Acinetobacter* spp were susceptible to tigecycline, <sup>(23)</sup> whereas Manoharan et al

(24) reported a low rate of susceptibility (42%) to tigecycline among multidrug resistant *Acinetobacter* spp. Various authors have reported a resistance rate to tigecycline varying from being nonexistent to 66% in *Acinetobacter* spp. A recent study from PGI, Chandigarh, India (30) reported 85.8% and 36% susceptibility to tigecycline among MDR *Acinetobacter* spp and carbapenem resistant MDR *Acinetobacter* spp isolates respectively. They reported very high such as 6 µg/ml & 32 µg/ml of MIC50 and MIC90 values of tigecycline against *Acinetobacter* spp. respectively. We too found higher MIC50 and MIC90 values of tigecycline like 1 µg/ml & 4 µg/ml respectively against *Acinetobacter* spp. Other studies from Singapore (27) & Thailand (31) also reported found higher MIC50 and MIC90 values of tigecycline against *Acinetobacter* spp.

In India, infections caused by Acb complex pose a therapeutic challenge owing to their multidrug resistance. (32) Many studies reported most of *Acinetobacter* isolates showed complete or high resistance to multiple drugs including Carbapenems. Neelam Taneja et al (30) reported that 41.5% of *Acinetobacter* spp isolated from complicated UTI were MDR and showed high resistance to cefotaxime (74.1%), gentamicin (79.5%), amikacin (73.2%), ciprofloxacin (72.8%), piperacillin-tazobactam (31.7%) and imipenem (25.4%).

In another study, (33) *Acinetobacter* spp isolated from blood stream infections showed very low susceptibility to many drugs like ceftazidime (44.6%), piperacillin-tazobactam (32.1%), amikacin (46.4%) and ciprofloxacin (48.2%). Whereas Karthika et al (34) reported that most of active isolates showed complete or high resistance to imipenem (100%), meropenem (89%), amikacin (80%), cefotaxime (89%) and ciprofloxacin (72%). We too found high resistance by *Acinetobacter* spp to multiple drugs similar to these studies.

Few antimicrobial agents remain that are active against a wide range of organisms. For gram negative organisms, carbapenems & polymyxins are highly active. However in present study, high resistance has been reported for many antimicrobials including carbapenems. In another study by Mezzatesta et al (35) from Italy had reported 90% of the isolates from their hospital to be resistant to first line drugs, with imipenem resistance being 50%.

## CONCLUSION

Thus tigecycline, with its ability to circumvent the common resistance mechanisms and its adequate microbiological activity against gram negative organisms, may make a welcome alternative for the treatment of multidrug resistant gram negative organisms.

## ACKNOWLEDGMENT

We are very thankful to Dr. Mrunal Patil, Dean and Dr. Hariprakash Gadde, Prof. & Head, Dept of Microbiology, for their timely support & valuable advice.

## REFERENCES

1. Falagas ME, Bliziotis IA. Pandrug-resistant Gram-negative bacteria: the dawn of the post-antibiotic era? *Int J Antimicrob Agents* 2007; 29: 630– 6.
2. Livermore DM, Woodford N. The  $\beta$ -lactamase threat in Enterobacteriaceae, *Pseudomonas* and *Acinetobacter*. *Trends Microbiol* 2006; 14: 413– 20.
3. Rossi F, Garcia P, Ronzon B, Curcio D, Dowzicky MJ. Rates of antimicrobial resistance in Latin America (2004-2007) and in vitro activity of the glycylicycline tigecycline and of other antibiotics. *Braz J Infect Dis* 2008; 12 (5):405-15.

4. Doan TL, Fung HB, Mehta D et al. Tigecycline: a glycylycylines antimicrobial agent. *Clin Ther* 2006; 28: 1079– 106.
5. Zhanel GG, Homenuik K, Nichol K et al. The glycylycylines: a comparative review with the tetracyclines. *Drugs* 2004; 64: 63– 88
6. Livermore DM. Tigecycline: what is it, and where should it be used? *J Antimicrob Chemother* 2005; 56: 611– 4.
7. Nathwani, D. 2005. Tigecycline: clinical evidence and formulary positioning *Int. J. Antimicrob. Agents* 25:185–192.
8. Pachon-Ibanez, M. E., M. E. Jimenez-Mejias, C. Pichardo, A. C. Llanos, and J. Pachon. 2004. Activity of tigecycline (GAR-936) against *Acinetobacter baumannii* strains, including those resistant to imipenem. *Antimicrob Agents Chemother.* 48:4479–4481.
9. Bradford, P. A. 2004. Tigecycline: a first in class glycylycylines. *Clin. Microbiol. News* 1.26:163–168.
10. Fritsche, T. R., H. S. Sader, M. G. Stilwell, M. J. Dowzicky, and R. N. Jones. 2005. Antimicrobial activity of tigecycline tested against organisms causing community-acquired respiratory tract infection and nosocomial pneumonia. *Diagn. Microbiol. Infect. Dis.* 52:187–193.
11. Fritsche, T. R., H. S. Sader, M. G. Stilwell, M. J. Dowzicky, and R. N. Jones. 2005. Potency and spectrum of tigecycline tested against an international collection of bacterial pathogens associated with skin and soft tissue infections (2000–2004). *Diagn. Microbiol. Infect. Dis.* 52:195–201
12. Kronvall, G., I. Karlsson, M. Walder, M. Sorberg, and L. E. Nilsson. 2006. Epidemiological MIC cut-off values for tigecycline calculated from E test MIC values using normalized resistance interpretation. *J. Antimicrob. Chemother.* 57:498–505
13. Ronald N. Jones, Mary Jane Ferraro, L. Barth Reller, Paul C. Schreckenberger, Jana M. Swenson, and Helio S. Sader. Multicenter Studies of Tigecycline Disk Diffusion Susceptibility Results for *Acinetobacter* spp. *Journal of Clinical Microbiology*; Jan. 2007:227–230.
14. Hoban, D. J., S. K. Bouchillon, B. M. Johnson, J. L. Johnson, and M. J. Dowzicky. In vitro activity of tigecycline against 6792 Gram-negative and Gram-positive clinical iso-lates from the global Tigecycline Evaluation and Surveillance Trial (TEST Program, 2004). *Diagn Microbiol Infect Dis* 2005; 52: 215– 27
15. Ruzin A, Keeney D, Bradford PA. AcrAB efflux pump plays a role in decreased susceptibility to tigecycline in *Morganella morganii* *Antimicrob Agents Chemother* 2005; 49:791– 3.
16. Falagas ME, Metaxas EI. Tigecycline for the treatment of patients with community-acquired pneumonia requiring hospitalization. *Expert Rev Anti Infect Ther* 2009; 7:913 -23
17. Winn W, Allen S, Janda W, Koneman E, Procop G, Schreckenberger P. the role of microbiology laboratory in the diagnosis of infectious diseases. guidelines to practice and management. Chapter 2. In: Koneman's Color Atlas and textbook of Diagnostic Microbiology. 6th

- Edn. Baltimore: Lippincott Williams and Wilkins 2006; 672-764.
18. Clinical and Laboratory Standards Institute (2012). Performance standards for antimicrobial susceptibility testing. 22<sup>nd</sup> Informational supplement. M100-S22. Wayne, PA: CLSI; 2012.
  19. Yong D, Lee K, Yum JH, Shin HB, Rossolini GM, Chong Y. Imipenem-EDTA disk method for differentiation of metallo- $\beta$ -lactamases-producing clinical isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *J Clin Microbiol* 2002; 40 : 3798-801
  20. Paterson DL. The epidemiological profile of infections with multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter* species. *Clin Infect Dis* 2006;43 (Suppl 2):S43-S48.
  21. Theodoros Kelesidis, Drosos E. Karageorgopoulos, Iosif Kelesidis and Matthew E. Falagas. Tigecycline for the treatment of multidrug-resistant Enterobacteriaceae: a systematic review of the evidence from microbiological and clinical studies. *Journal of Antimicrobial Chemotherapy* (2008) 62, 895–904
  22. Mary L Townsend, Melanie W Pound, Richard H Drew. Tigecycline in the treatment of complicated intra-abdominal and complicated skin and skin structure infections *Therapeutics and Clinical Risk Management* 2007;3(6) 1059–1070
  23. Behera B, Das A, Mathur P, kapil A, Gadepalli R, Dhawan B. Tigecycline susceptibility report from an Indian tertiary care hospital. *Indian J Med Res* 2009;129:446-50
  24. Manoharan A, Chatterjee S, Madhan S, Mathai D. evaluation of tigecycline activity in clinical isolates among Indian medical centers. *Indian J Pathol Microbiol* 2010;53 (4):734-37
  25. Shanthi M., Uma Sekar. In vitro Activity of Tigecycline against Gram Positive and Gram Negative Isolates in a Tertiary Care Hospital. *Journal of Clinical and Diagnostic Research*. 2011 December, Vol-5(8): 1559-1563.
  26. Souli M, Kontopidou FV, Koratzanis E, Antoniadou A, Giannitsioti E, Evangelopoulou P, et al. In vitro activity of tigecycline against multiple-drug-resistant, including pan-resistant, gram-negative and gram positive clinical isolates from Greek hospitals. *Antimicrob Agents Chemother* 2006; 50: 3166-3169.
  27. Thean-Yen Tan, Lily SY Ng. Susceptibility of Multi-resistant Gram-negative Bacilli in Singapore to Tigecycline as Tested by Agar Dilution. *Ann Acad Med Singapore* 2007; 36: 807-10.
  28. Carmen Betriu , Iciar Rodríguez - Avial, Blas Ali Sánchez , María Gómez, Juan Álvarez, Juan J. Picazo, and Spanish Group of Tigecycline†. In Vitro Activities of Tigecycline (GAR-936) against Recently Isolated Clinical Bacteria in Spain. *Antimicrobial Agents and Chemotherapy*, Mar. 2002; 46 (3): 892–895.
  29. Ralf Rene Reinert, Donald E. Low, Flavia Rossi, Xiaojiang Zhang, Chand Wattal and Michael J. Dowzicky. Antimicrobial susceptibility among organisms from the Asia/Pacific Rim, Europe and Latin and North America collected as part of TEST and the in vitro activity of tigecycline. *Journal of Antimicrobial Chemotherapy*. 2007; 60: 1018–1029.

30. Neelam Taneja, Gagandeep Singh, Meenakshi Singh & Meera Sharma. Emergence of tigecycline & colistin resistant *Acinetobacter baumannii* in patients with complicated urinary tract infections in north India. *Indian J Med Res.* 2011; 133: 681-684.
31. Surapee Tiengrim ,Chanwit Tribuddharat MD, Visanu Thamlitkul MD. In Vitro Activity of Tigecycline against Clinical Isolates of Multidrug-Resistant *Acinetobacter baumannii* in Siriraj Hospital, Thailand. *J Med Assoc Thai* 2006; 89 (Suppl 5): S102-5
32. Azim A, Dwivedi M, Rao PB, Baronia AK, Singh RK, Prasad KN, et al. Epidemiology of bacterial colonization at intensive care unit admission with emphasis on extended-spectrum beta-lactamases and metallo-beta-lactamase producing Gram-negative bacteria - an Indian experience. *J Med Microbiol* 2010; 59: 955-60.
33. Prabhash K, Medhekar A, Ghadyalpatil N, Noronha V, Biswas S, Kurkure P, et al. . Blood stream infections in cancer patients: a single center experience of isolates and sensitivity pattern. *Indian J Cancer* 2010; 47: 184-8.
34. Uma Karthika R, Srinivasa Rao R, Sahoo S, Shashikala P, Kanungo R, Jayachandran S, et al. . Phenotypic and genotypic assays for detecting the prevalence of metallo-beta-lactamases in clinical isolates of *Acinetobacter baumannii* from a South Indian tertiary care hospital. *J Med Microbiol.* 2009; 58: 430-5.
35. Mezzatesta M I, Trovato G, Gona F, Nicolosi VM, Nicolosi D, Carattoli A, et al. In vitro activity of tigecycline and comparators against carbapenem-susceptible and resistant *Acinetobacter baumannii* clinical isolates in Italy. *Ann Clin Microbiol Antimicrob* 2008; 7: 4.

How to cite this article: Mane M, Gangurde N, Phatale S. In vitro activity of tigecycline against multidrug resistant gram negative bacilli as evaluated by disc diffusion method and E-test. *Int J Health Sci Res.* 2014;4(1):85-92.

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